

PEJABAT TIMBALAN KETUA PENGARAH KESIHATAN (PERUBATAN) KEMENTERIAN KESIHATAN MALAYSIA ARAS 7, BLOK E1, KOMPLEKS E PUSAT PENTADBIRAN KERAJAAN PERSEKUTUAN 62590 PUTRAJAYA



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SEPERTI SENARAI EDARAN

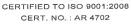
YBhg. Datuk/Dato'/Datin/Tuan/Puan,

PENAMBAHBAIKAN "GUIDELINE ON STANDARDIZATION OF WORKLOAD DATA COLLECTION 4TH EDITION 2016"

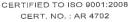
Dengan segala hormatnya saya merujuk kepada perkara di atas dan surat rujukan KKM.600-27/4/2 (15) bertarikh 30 Jun 2016 adalah berkaitan.

- 2. Untuk makluman YBhg. Datuk/Datoʻ/Datin/Tuan/Puan, Bengkel Beban Kerja Patologi telah pun berjaya diadakan pada 22 Ogos 2016 di Institut Kanser Negara, Putrajaya. Bengkel ini telah dihadiri oleh wakil-wakil makmal Patologi hospital-hospital KKM, makmal-makmal Kesihatan Awam dan juga Institusi-Institusi KKM seperti Pusat Darah Negara dan Institut Penyelidikan Perubatan.
- 3. Susulan daripada bengkel tersebut, terdapat beberapa penambahbaikan telah dilakukan terhadap garis panduan ini. Sehubungan dengan itu, diharapkan garis panduan yang telah ditambah baik ini dapat digunapakai oleh semua makmal di hospital, kemudahan kesihatan awam dan institusi lain yang berkaitan dalam Kementerian Kesihatan Malaysia (KKM).
- 4. Garis panduan yang telah ditambah baik ini boleh didapati di laman sesawang Perkhidmatan Patologi (www.patologi.gov.my) dan laman sesawang KKM bagi rujukan YBhg. Datuk/Datoʻ/Datin/Tuan/Puan. Sebarang pertanyaan mengenai garis panduan tersebut boleh diajukan kepada YBhg. Datin Dr. Nik Noraihan binti Nik Mustapha, Hospital Sultanah Bahiyah, Kedah di talian 04-740 6799 / 6774 atau nikraihan@kdh.moh.gov.my.











CERTIFIED TO ISO 9001:2008 CERT, NO. : AR 4702

Segala kerjasama YBhg. Datuk/Dato'/Datin/Tuan/Puan di dahulukan dengan 5. ucapan terima kasih.

Sekian.

"BERKHIDMAT UNTUK NEGARA"

Saya yang menurut perintah,

DR. Ariveru KAZID BIN SALLEH (MMC:28566) Tengarah Amelian Perubatan Bahagian Amelian Perubatan Kementarian Keshatan Malaysia

(DATUK DR. JEYAINDRAN TAN SRI SINNADURAI)

Timbalan Ketua Pengarah Kesihatan (Perubatan)

Kementerian Kesihatan Malaysia

s.k.:

Ketua Pengarah Kesihatan Kementerian Kesihatan Malaysia

Ketua Perkhidmatan Patologi Kebangsaan

SENARAI EDARAN

Pengarah Jabatan Kesihatan Negeri Perlis

Pengarah Jabatan Kesihatan Negeri Kedah

Pengarah Jabatan Kesihatan Negeri Pulau Pinang

Pengarah Jabatan Kesihatan Negeri Perak

Pengarah Jabatan Kesihatan Wilayah Persekutuan Kuala Lumpur / Putrajaya

Pengarah Jabatan Kesihatan Negeri Selangor

Pengarah Jabatan Kesihatan Negeri Negeri Sembilan

Pengarah Jabatan Kesihatan Negeri Melaka

Pengarah Jabatan Kesihatan Negeri Johor

Pengarah Jabatan Kesihatan Negeri Pahang

Pengarah Jabatan Kesihatan Negeri Terengganu

Pengarah Jabatan Kesihatan Negeri Kelantan

Pengarah Jabatan Kesihatan Negeri Sarawak

Pengarah Jabatan Kesihatan Negeri Sabah

Pengarah Jabatan Kesihatan Wilayah Persekutuan Labuan

Pengarah Hospital Kuala Lumpur

Pengarah Institut Kanser Negara

Pengarah Pusat Darah Negara

Pengarah Institut Perubatan Respiratori

Pengarah Institut Penyelidikan Perubatan

Pengarah Makmal Kesihatan Awam Kebangsaan

Pengarah Makmal Kesihatan Awam Ipoh

Pengarah Makmal Kesihatan Awam Johor Bahru

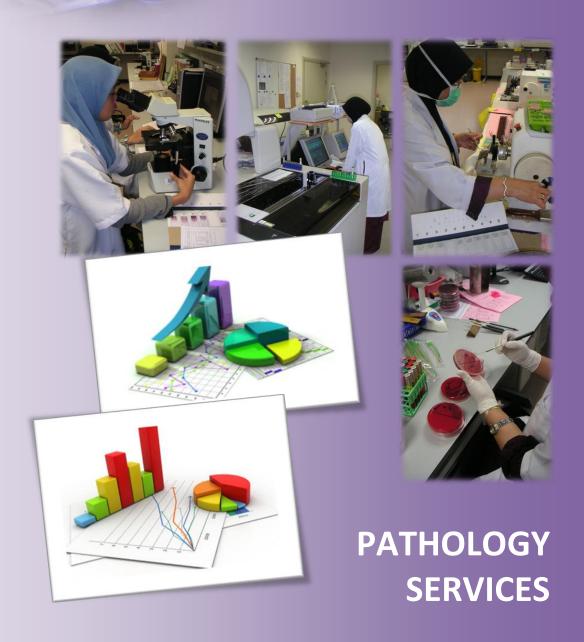
Pengarah Makmal Kesihatan Awam Kota Bharu

Pengarah Makmal Kesihatan Awam Kota Kinabalu

Timbalan Pengarah Pusat Informatik Kesihatan



GUIDELINE ON STANDARDIZATION OF WORKLOAD DATA COLLECTION 4th EDITION 2016



MINISTRY OF HEALTH
MALAYSIA

GUIDELINE ON STANDARDIZATION OF WORKLOAD DATA COLLECTION $\mathbf{4}^{\text{th}}$ EDITION - 2016

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1. INTRODUCTION

Workload of Pathology services forms the basis to assist Ministry of Health (MOH) Malaysia in planning for resource management (e.g. manpower, budget and equipment), policy decisions, as well as monitoring the utilization of pathology services.

The first guideline on workload calculation and data collection was published in 1999. A few discrepancies in and alterations to the first edition led to issuance of the 2nd and 3rd editions in quick succession (August 2000 and January 2002, respectively). The 3rd edition guideline has since been used by laboratories in Ministry of Health (MOH) hospitals and health clinics. Since then, there has been marked progression in the range of tests offered, as well as laboratory testing methodologies, especially from manual to automation. In 2009, an attempt was made to revise and update the guideline in data collection. However, the draft of the revised guideline that was produced then did not get to be endorsed due to technical issues.

In 2014, an initiative was again made to revise and update the existing guideline and the process continued through in 2015. An initial meeting was held in August 2014 to discuss on this matter, which was chaired by the Head of Pathology Services and attended by representatives from individual Pathology disciplines from hospital laboratory services, as well as from Institute for Medical Research (IMR), National Blood Centre (*Pusat Darah Negara*, PDN), National Public Health Laboratory (*Makmal Kesihatan Awam Kebangsaan*, MKAK), *Bahagian Pembangunan Kesihatan Keluarga* (BPKK) *and Pusat Informatik Kesihatan* (PIK). It was agreeable during this meeting that the draft document on the revised guideline on workload data collection, which was produced in 2009 needed further updating due to expansion of test lists and methodologies. In addition, the format of workload submission also needed to be revised to allow more detailed, representative and informative workload reporting.

Finally, the current updated guideline was developed to ensure that data collection is conducted in a standardised manner by all laboratories in hospitals, public health facilities and other relevant institutions within MOH.

2. REVIEW OF DATA COLLECTION PRINCIPLES AND REPORTING FORMAT

Workload is represented by number of tests performed, as defined by and unique to an individual discipline. Data generated should be logical, meaningful and useful for a particular discipline. In certain laboratories/pathology disciplines, extensive automation has taken place, whilst in others, some components of manual testing are still indispensible. Method of data calculation and collection for one particular discipline should not be literally compared to another discipline, especially in terms of number of tests generated. As much as possible, multiplication factor is avoided in calculating number of tests to prevent potential irregularities in data collection. Data collected should, among others, allow some reflection on the resources (namely budget and manpower) needed in running an individual laboratory. To assist that, in addition to number of tests, data are also collected in the forms of group of tests and number of specimens.

Standardised discipline-specific soft copy excel forms (*Borang Beban Kerja Bulanan* 1/2016 - *Patologi Anatomik / Hematologi / Patologi Kimia / Mikrobiologi Perubatan*) for granular workload data collection had been prepared to ease the data collection, calculation and submission process. National Pathology Workload Committee needs to be directly informed to fascilitate addition of tests that are not yet listed and need to be included in the test lists.

To accommodate the changes in the format of workload data collection, additional discipline-specific forms, Borang Ringkasan Beban Kerja 1/2016 - Patologi Anatomik / Hematologi / Patologi Kimia / Mikrobiologi Perubatan, had also been created to enhance workload data reporting within the Pathology services. These forms allow data to be recorded according to test groups. It is also in excel format and is auto-filled, in tandem with data collection/recording activity using discipline-specific Borang Beban Kerja Patologi Bulanan 1/2016. The PER-SS 206 – Pin 1/2000 form will continue to be used for monthly workload reporting to MOH.

3. SCOPE OF DOCUMENT

This guideline is applicable for use by diagnostic laboratories dealing with human samples, within the Ministry of Health, Malaysia. Such laboratories include:

- 3.1 Hospital laboratories
- 3.2 Health Clinic laboratories
- 3.3 Institutes and agencies, which include:
 - 3.3.1. Institute for Medical Research (IMR)
 - 3.3.2. National Blood Bank Centre (Pusat Darah Negara, PDN)
 - 3.3.3. National Cancer Institute (Institut Kanser Negara, IKN)
 - 3.3.4. Institute of Respiratory Medicine (Institut Perubatan Respiratori, IPR)
 - 3.3.5. Women and Children Hospitals (WCH)
- 3.4 Public Health Laboratories:
 - 3.4.1 National Public Health Laboratory (Makmal Kesihatan Awam Kebangsaan, MKAK)
 - 3.4.2 Regional Public Health Laboratories (Makmal Kesihatan Awam, MKA)

4. NATIONAL PATHOLOGY WORKLOAD COMMITTEE

- 4.1 Data collection and minding is under the perview of National Pathology Workload Committee (*Jawatankuasa Beban Kerja Patologi Kebangsaan*).
- 4.2 Data collection and minding activities include but not limited to:
 - 4.2.1 Six-monthly compilation of granular and aggregate workload data, which are received from state pathologists, institutes and agencies, as well as MKAK.
 - 4.2.2 Annual reporting on national pathology workload data. This is to aid the National Head of Pathology Services (*Ketua Perkhidmatan Patologi Kebangsaan*) and Heads of Pathology Disciplines in making annual report on pathology services, as well as for purpose of continuous service and activity planning.

- 4.3 The committee consists of:
 - 4.3.1 Chairman (Pathologist)
 - 4.3.2 Vice Chairman (Pathologist)
 - 4.3.3 Representatives from each pathology discipline i.e. Anatomic Pathology, Chemical Pathology, Haematology and Medical Microbiology
 - 4.3.4 Representatives from institutes, agencies and departmental laboratories, which include IMR, PDN, IPR, IKN and Molecular and Cytogenetic Laboratory of Hospital Kuala Lumpur.
 - 4.3.5 Representatives from Bahagian Pembangunan Kesihatan Keluarga (BPKK).
 - 4.3.6 Representatives from National and Regional Public Health Laboratories (MKAK and MKA).

5. GENERAL PRINCIPLES OF DATA COLLECTION

- 5.1 Only tests performed by individual laboratories are considered as their true workload.
- 5.2 Data on outsourced and referred tests are to be separately collected and recorded. This will give some information on the pre-analytical manpower involved in specimen preparation prior to sending out to external laboratories. Workload on outsourced and referred tests will also be useful in policy making and planning for centralization or decentralization of services.

Note:

- Referred samples/tests Apply to samples sent to MOH laboratories or other centres, which do not incur extra cost to the primary/referring laboratory.
- ii. Outsourced samples/tests Apply to samples sent to private laboratories or other centres and the primary/outsourcing laboratories are charged for the services rendered.
- 5.3 Only tests performed on patient's specimens are included in the workload. Internal Quality Control (IQC) and External Quality Assurance (EQA) tests are excluded.
- 5.4 There may be some tests that are performed by more than one discipline in different hospital settings, either by tradition or by default of where the instrument/analyser is placed. Examples of such tests include CRP, C3, C4, IgG, IgA and IgM, which are done either in Chemical Pathology or Serology/Immunology laboratories. In principle, "ownership" of tests by individual discipline should take into consideration the technical and consultancy accountability.
- 5.5 Clerical work (e.g demography and typing) and result validation are not included in workload collection.
- 5.6 Workload on 'non-test' technical activities may be separately collected and recorded to allow some reflection on pre-analytical manpower and other resources involved. Such

activities include media preparation for microbiological culture and cytogenetic testing. Recording of this workload utilises a separate excel sheet in individual discipline's *Borang Beban Kerja Patologi Bulanan* and *Borang Beban Kerja Patologi Negeri*. Data generated is not included as part of workload submission to MOH.

- 5.7. The personnel responsible for providing data on workload may either be a Science Officer or a Medical Laboratory Technologist (MLT) in charge. Head of Department/Officer in charge is responsible to verify the generated data.
- 5.8. All MOH laboratories are to report their monthly aggregate workload data to Ministry of Health using *PER-SS 206 (Pin. 1/2000)* form, through existing channels (see section on preparation and submission of reports below).
- 5.9 All hospital laboratories are also to submit detailed, as well as summarised workload data to their respective State Pathologists on monthly basis. The State Pathologists will in turn, compile these data and submit them biannually to the National Pathology Workload Committee (see section on preparation and submission of reports below).
- 5.10 State Pathologist is responsible to verify and monitor data sent to MOH from all laboratories within the state.

6. PREPARATION & SUBMISSION OF REPORTS

6.1 Monthly Reports

- 6.1.1 Verified workload data shall be submitted to Ministry of Health by 15th day of the subsequent month, using *PER-SS 206 (Pin. 1/2000)* form, through existing channels (refer Diagram 1 on Workload Data Submission).
- 6.1.2 *Jabatan Kesihatan Negeri* shall compile and submit workload data for the state to Ministry of Health, also using *PER-SS 206 (Pin. 1/2000)* form.
- 6.1.3 In addition, hospital laboratories shall submit workload data in soft copy to State Pathologists, by 15th day of subsequent month using the following forms:
 - i. Discipline-specific Borang Beban Kerja Patologi Bulanan 1/2016
 - ii. Discipline-specific Borang Ringkasan Beban Kerja Patologi 1/2016

6.2 Biannual Reports

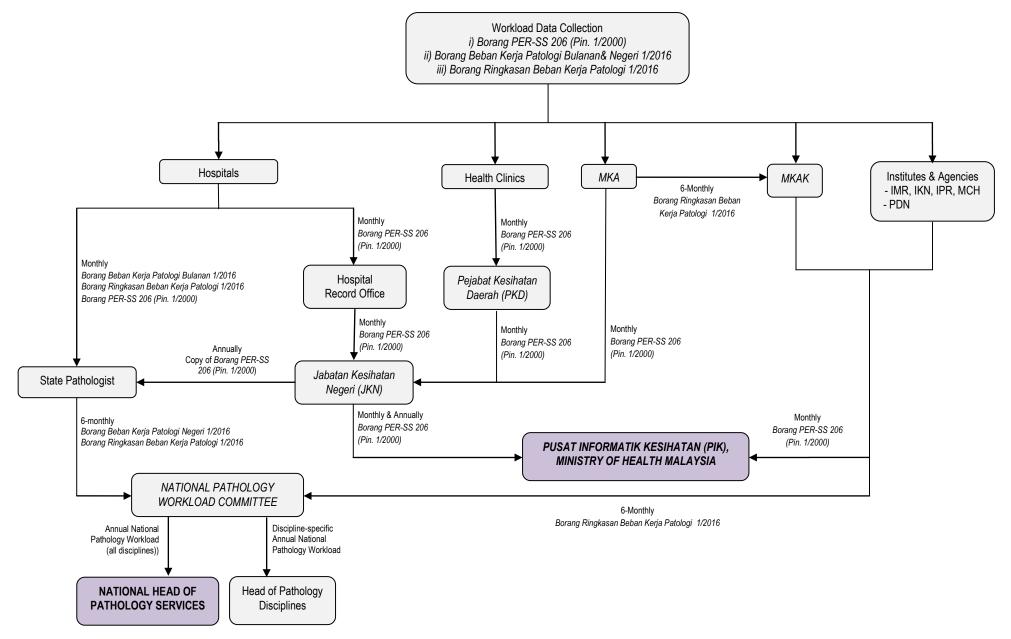
6.2.1 State Pathologists shall submit 6-monthly workload data for all hospitals in their states, using discipline-specific *Borang Beban Kerja Patologi Negeri 1/2016* and *Borang Ringkasan Beban Kerja Patologi 1/2016* to National Pathology Workload

- Committee, by 30th July for January June data and 31st January of the following year for January December data.
- 6.2.2 Institutes and agencies, as well as, *MKAK* shall submit their workload data, using *Borang Ringkasan Beban Kerja Patologi* 1/2016 to National Pathology Workload Committee by 30th July for January June data and 31st January of the following year for January December data.
- 6.2.3 *MKA* shall submit their workload data to *MKAK* using *Borang Ringkasan Beban Kerja Patologi 1/2016* by 30th July for January June data and 31st January of the following year for January December data.

6.3 Annual Reports

- 6.3.1 *Jabatan Kesihatan Negeri* shall compile and submit annual workload data for the state to PIK, Ministry of Health, also using *PER-SS 206 (Pin. 1/2000)* form. In addition, a copy shall be given to State Pathologist.
- 6.3.2 National Pathology Workload Committee shall compile workloads submitted by State Pathologists, Institutes, Agencies and MKAK into Annual National Pathology Workload. Information shall be made available to the National Head of Pathology Services (*Ketua Perkhidmatan Patologi Kebangsaan*).
- 6.3.3 Detailed workload data for individual discipline (for hospital laboratories) will also be given to the respective Head of Disciplines.
- 6.3.4 Formal requests for data by other parties will be entertained subject to approval by the National Head of Pathology Services, and where appropriate, by the Director General of Health.

DIAGRAM 1: WORKLOAD DATA SUBMISSION



PER-SS 206 (Pin. 1/2000)

HEALTH MANAGEMENT INFORMATION SYSTEM MINISTRY OF HEALTH, MALAYSIA REPORT OF LABORATORY WORKLOAD: IN-PATIENT / OUT-PATIENT

FOR THE MONTH:______ YEAR :_____

Location of General, District Hospital, Institutions and Public Health Facilities	Biochemistry	Microbiology	Haematology	Histopathology	Cytology	Forensic	Total	Level of Laboratory
1	2	3	4	5	6	7	8	9
TOTAL								

Tanda Tangan Ketua Jabatan

7. CALCULATION AND RECORDING OF WORKLOAD DATA

7.1 ANATOMICAL PATHOLOGY

- 7.1.1 Workload data collection is assisted by using several pre-prepared worksheets (see subsequent section).
- 7.1.2 Workload reporting in Anatomical Pathology involves:
 - 7.1.2.1 Monthly submission of aggregate Histopathology and Cytopathology workload data to MOH, via existing channel, using *Borang PER-SS 206* (*Pin. 1/2000*).
 - 7.1.2.2 Monthly submission of detailed (granular), as well as, summarised Histopathology and Cytopathology workload data to State Pathologist, using soft copy 'Borang Beban Kerja Patologi Bulanan 1/2016-Patologi Anatomik' and 'Borang Ringkasan Beban Kerja Patologi 1/2016- Patologi Anatomik', respectively.
 - 7.1.2.3 Six-monthly submission of specimen complexity data to State Pathologist using additional pre-prepared soft copy sheet, as part of 'Borang Beban Kerja Patologi Bulanan 1/2016-Patologi Anatomik'.
 - 7.1.2.4 Six-monthly submission of data on individual pathologist's workload to State Pathologist using additional pre-prepared soft copy sheet, also as part of 'Borang Beban Kerja Patologi Bulanan 1/2016-Patologi Anatomik'.
- 7.1.3 State pathologists are to biannually submit data on detailed Histopathology and Cytopathology workload, specimen complexity and individual pathologist's workload to National Pathology Workload Committee, using 'Borang Beban Kerja Patologi Negeri 1/2016 Patologi Anatomik'.
- 7.1.4 Categorisation of specimen complexity shall be based on 'Histopathology Specimen Complexity Guide' provided in this guideline. Laboratories are advised to use own clinical discretion/judgement in categorising specimens that may not be listed.
- 7.1.5 Individual pathologist's workload shall take into account cases that are co-reported/co-verified by more than one specialist.
- 7.1.6 A case is defined as one or more specimens belonging to one patient that is/are registered and given one unique laboratory identifier number. Each case registered is counted as one. If there are more than one specimen received for one case, each specimen is counted as one and the total number of specimens is separately recorded.
- 7.1.7 Second opinion (referral) cases are those that are received for consultation, including those requested by local clinicians (e.g. for review by in-house pathologists prior to initiation of chemotherapy). Workload for cases that are received for special or

- immunohistochemical staining assistance are counted under "Specialised tests" section.
- 7.1.8 If only pre-stained slides are received for cases for second opinion, workload is captured by number of cases only. If paraffin blocks are received and additional sections and stains are performed, number of H&E slides and special/immunohistochemical stains (if applicable) are recorded but not the number of blocks.
- 7.1.9 Intraoperative frozen sections (FS) are fozen section assessments that are done for purpose of either assissting intraoperative surgical management or transplant organ procurement. Frozen sections performed as part of complex biopsies (e.g. renal, skin and muscle biopsies) are not included.
 - 7.1.9.1 Count all paraffin blocks and and H&E slides produced for each FS case (including subsequent additional sampling done for the same specimen).
 - 7.1.9.2 If the laboratory subsequently receives additional surgical specimen(s) for the same patient, workload for the new specimen(s) is recorded/counted under 'Routine Surgical Pathology'.
- 7.1.10 In most centres, trephine biopsies are read by haematologists, together with bone marrow aspirates. However, if they are separately read by Anatomic Pathologists, then the number of cases and specimens are also to be recorded.
- 7.1.11 For autopsy cases, multiple organs/tissues are frequently collected in one container. Therefore, number of specimens are not counted. For centres which forensic autopsy slides are read by anatomic pathologists, number of cases are to be recorded. If read by forensic pathologists, then only number of paraffin blocks and H&E slides are counted as laboratory workload. Clinical autopsies performed are recorded in individual pathologists workload section.
- 7.1.12 For oral pathology cases that are reported by anatomic pathologists, they are to be recorded under routine surgical pathology and their complexity classified accordingly. Oral pathology cases that are reported by dental pathologists are separately recorded for purpose of capturing laboratory tissue processing and H&E slide preparation workload.
- 7.1.13 For specialised tests e.g. immunohistochemical stains, only number of test slides are captured. Control slides (separately stained from test tissue) are not counted. This is to ensure more accurate workload data collection, as well as to encourage better laboratory management.
- 7.1.14 The number of cell blocks produced for FNA and Non-gynaecological specimens is captured under Cytology workload, even though processing is done in Histopathology laboratory.

- 7.1.15 Seminal fluid analysis as part of investigation for fertility, is considered as a cytology test, even though traditionally, it may have been performed in microbiology laboratory. Medicolegal samples, including vaginal swabs such as in rape cases, are not handled by diagnostic cytology laboratory.
- 7.1.16 For test category that may not be listed, workload submission is by temporarily adding it under 'others'. Please inform National Pathology Workload Committee via Head of Discipline, of the test name/category that needs to be added into the test list in both Borang Beban Kerja Patologi Bulanan 1/2016 Patologi Anatomik and Borang Beban Kerja Patologi Negeri 1/2016 Patologi Anatomik.
- 7.1.17 Workload calculation and recording: Refer to worksheets and corresponding forms on page 12 till page 22.

PATHOLOGY SERVICES, MINISTRY OF HEALTH MALAYSIA WORKSHEET FOR ANATOMIC PATHOLOGY WORKLOAD (HISTOPATHOLOGY) 1/2016 – For Laboratory Use Only

Hospital:	_Month:	Year:	Reported by:

TESTS PERFORMED BY INDIVIDUAL LABORATORY

No.	Test	Number of cases	Number of specimens	No. of blocks	No. of H&E slides	No. of Tests	GRAND TOTAL
1	Routine Surgical Pathology					NA	NA
2	Received for 2 nd opinion (Referred in)					NA	NA
3	Intraoperative Frozen section					NA	NA
4	Trephine biopsy	NA if reported by Haematologist	NA if reported by Haematologist			NA	NA
5	Autopsy (Clinical and Forensic)	NA if reported by Forensic pathologist	NA			NA	NA
6	Oral pathology (cases reported by dental pathologists)	NA	NA			NA	NA
7	Others (if test/category not listed)					NA	NA
	SPECIALISED TESTS						
8	Immunohistochemical stain	NA	NA	NA	NA		
9	Special stain	NA	NA	NA	NA		
10	Enzyme histochemical stain	NA	NA	NA	NA		
11	Immunofluorescence stain	NA	NA	NA	NA		
12	Others (if test/category not listed)	NA	NA	NA	NA		
	DIAGNOSTIC MOLECULAR TESTS						
13	Real time- Polymerase Chain Reaction (RT-PCR)	NA	NA	NA	NA		
14	Fluorescence In Situ Hybridisation (FISH)	NA	NA	NA	NA		
15	Dual/Chromogenic In Situ Hybridisation (DISH/CISH)	NA	NA	NA	NA]
16	Others (if test/category not listed)	NA	NA	NA	NA		
	TOTAL	NA	NA			NA	NA
	GRAND TOTAL					NA	NA

NA – Not applicable

PATHOLOGY SERVICES, MINISTRY OF HEALTH MALAYSIA WORKSHEET FOR ANATOMIC PATHOLOGY WORKLOAD (HISTOPATHOLOGY) 1/2016 – For Laboratory Use Only

RRED TESTS	(specimens/samples sent to other labora	tories for testing and are NON-CHARGEABLE)	
0.	Test	Referral Centre	Total No. of Cases
1			
2			
3			
	TOTAL		

No.	Test	Outsource Centre	Total No. of Cases
1			
2			
3			
	TOTAL		

EXPLANATION FOR WORKSHEET FOR ANATOMIC PATHOLOGY WORKLOAD (HISTOPATHOLOGY) PATHOLOGY SERVICES, MINISTRY OF HEALTH MALAYSIA WORKSHEET FOR ANATOMIC PATHOLOGY WORKLOAD (HISTOPATHOLOGY) 1/2016 – For Laboratory Use Only

Hospital:	Month:	Year:	Reported by:
			' '

TESTS PERFORMED BY INDIVIDUAL LABORATORY

No.	Test	Number of cases	Number of specimens	No. of paraffin blocks	No. of H&E slides	TOTAL No. of Tests	GRAND TOTAL
1	Routine Surgical Pathology	No. of cases received	No. of specimens received	No. of blocks	No. of slides	NA	NA
	100000000000000000000000000000000000000	and registered	& examined for each case No .of specimens received	prepared No. of blocks	prepared No. of slides	,	
2	Received for 2 nd opinion (Referred in)	No. of cases received	& examined for each case	prepared	prepared	NA	NA
_	necessaries 2 opinion (necessarily	and registered	(if applicable)	(if applicable)	(if applicable)		
3	Intraoperative Frozen section	No. of cases received	No .of specimens received	No. of blocks	No. of slides	NA	NA
		and registered NA if reported by	& examined for each case NA if reported by	prepared No. of blocks	prepared No. of slides		
4	Trephine biopsy	Haematologist	Haematologist	prepared	prepared	NA	NA
5	Autopsy (Clinical and Forensic)	NA if reported by	NA .	No. of blocks	No. of slides	NA	NA
	Autopsy (Clinical and Forensic)	Forensic pathologist	IVA	prepared	prepared	IVA	NA
6	Oral pathology (cases reported by dental pathologists)	NA	NA	No. of blocks	No. of slides	NA	NA
		No. of cases received	No .of specimens received	prepared No. of blocks	prepared No. of slides		
7	Others (if test/category not listed)	and registered	& examined for each case	prepared	prepared	NA	NA
	SPECIALISED TESTS						
8	Immunohistochemical stain	NA	NA	NA	NA	No. of stains performed	
9	Special stain	NA	NA	NA	NA	No. of stains performed	(Aggregate no of
10	Enzyme histochemical stain	NA	NA	NA	NA	No. of stains performed	stains performed
11	Immunofluorescence stain	NA	NA	NA	NA	No. of stains performed	for 8, 9,10,11,12)
12	Others (if test/category not listed)	NA	NA	NA	NA	No. of stains performed	
	DIAGNOSTIC MOLECULAR TESTS						
13	Real time- Polymerase Chain Reaction (RT-PCR)	NA	NA	NA	NA	No. of tests performed	
14	Fluorescence In Situ Hybridisation (FISH)	NA	NA	NA	NA	No. of tests performed	Aggregate no of
15	Dual/Chromogenic In Situ Hybridisation (DISH/CISH)	NA	NA	NA	NA	No. of tests performed	tests performed for 13,14,15,16)
16	Others (if test/category not listed)	NA	NA	NA	NA	No. of tests performed	
	TOTAL	NA	NA	Total no. of paraffin blocks	Total no. of H&E slides	NA	NA
	GRAND TOTAL	Total no. of cases	Total no. of specimens	Aggregate no. of b	locks & H&E slides	NA	NA

NA – Not applicable

PATHOLOGY SERVICES, MINISTRY OF HEALTH MALAYSIA WORKSHEET FOR ANATOMIC PATHOLOGY WORKLOAD (CYTOLOGY) 1/2016 – For Laboratory Use Only

Hospital: ______ Month: _____ Year: _____ Reported by: _____

TESTS	PERFORMED BY INDIVIDUAL LABORATORY				
No.	Test	No. of cases	No. of specimens	No. of slides	No. of cell blocks
1	Gynaecology conventional				NA
2	Gynaecology liquid base				NA
3	Non-gynaecology				
4	Fine Needle Aspiration				
5	Others (if test/ category not listed)				
	TOTAL	NA	NA		
	GRAND TOTAL				
REFE No.	RRED TESTS (specimens/samples sent to other la Test		ION-CHARGEABLE)	Total No	o. of Cases
1					
2					
3	TOTAL				
OUTS	OURCED TESTS (specimens/samples sent to other	er laboratories for testing and a	re CHARGEABLE)		
No.	Test	Outsour	ce Centre	Total No	o. of Cases
1					
2					
3	TOTAL				
					Page 1 of 1
					.00 = 0.1

EXPLANATION FOR WORKSHEET FOR ANATOMIC PATHOLOGY WORKLOAD (CYTOLOGY) PATHOLOGY SERVICES, MINISTRY OF HEALTH MALAYSIA WORKSHEET FOR ANATOMIC PATHOLOGY WORKLOAD (CYTOLOGY) 1/2016 – For Laboratory Use Only

	Hospital:		Month: Year:	Reported by:	
TESTS	PERFORMED BY INDIVIDUAL LABORATORY				
No.	Test	No. of cases	No. of specimens	No. of slides	No. of cell blocks
1	Gynaecology conventional	No. of cases registered	As for no. of cases	No. of slides prepared	NA
2	Gynaecology liquid base	No. of cases registered	As for no. of cases	No. of slides prepared	NA
3	Non-gynaecology	No. of cases registered	No. of specimens received for each case	No. of slides prepared	No. of blocks prepared
4	Fine Needle Aspiration	No. of cases registered	No. of specimens received for each case	No. of slides prepared	No. of blocks prepared
5	Others (if test/category not listed)	No. of cases registered	No. of specimens received for each case	No. of slides prepared	No. of blocks prepared
	TOTAL	NA	NA	Total no. of slides prepared	Total no. of blocks prepared
	GRAND TOTAL	Total no. of cases	Total no. of specimens	Total no. of tests (total	no. of slides & cell blocks)
REFER	RRED TESTS (specimens/samples sent to other	laboratories for testing and are	NON-CHARGEABLE)		
No.	Test	<u> </u>	Referral Centre	Total No	o. of Cases
1					
2					
3					
	TOTAL				
OUTS	OURCED TESTS (specimens/samples sent to ot	ther laboratories for testing and	are CHARGEABLE)		
No.	Test		Outsource Centre	Total No	o. of Cases
1					
2					
3					
	TOTAL				

PATHOLOGY SERVICES MINISTRY OF HEALTH, MALAYSIA

REPORT ON LABORATORY WORKLOAD: ANATOMIC PATHOLOGY

FOR THE OF MONTH:	YEAR :

		HISTOPATHOLOGY						CYTOPATHOLOGY		
Month or Location of Hospitals, Institutions and Public Health Facilities	No. of Cases	No. of H&E Slides & Tissue Blocks (a)	No. of Specialised Tests (b)	No. of Diagnostic Molecular Tests (c)	Total No. of Specimens	Total No. of Tests (a + b + c)	No. of Cases	Total No. of Specimens	Total No. of Tests	

PATHOLOGY SERVICES, MINISTRY OF HEALTH MALAYSIA WORKSHEET FOR ANATOMIC PATHOLOGY WORKLOAD (HISTOPATHOLOGY SPECIMEN COMPLEXITY) 1/2016 – For Laboratory Use Only

	JAN - JUN Year:
	JAN - DIS Year:
Hospital:	 Reported by:

No.	Specimen Category	Number of cases	Number of specimens
1	Simple	X No. of cases received and registered	X No. of specimens received & examined for each case
2	Medium	X No. of cases received and registered	X No. of specimens received & examined for each case
3	Complex	X No. of cases received and registered	X No. of specimens received & examined for each case
4	Very complex	X No. of cases received and registered	X No. of specimens received & examined for each case
5	Complex biopsies & Referral cases	X No. of cases received and registered	X No. of specimens received & examined for each case
	TOTAL		

PATHOLOGY SERVICES, MINISTRY OF HEALTH MALAYSIA HISTOPATHOLOGY SPECIMEN COMPLEXITY GUIDE 1/2016 – For Laboratory Use Only

Kindly use your clinical discretion/judgement in categorising specimens that may not be listed here

Simple	Medium	Complex	Very Complex	Complex biopsies & Referral cases
Non-complex excision/ small specimen:	a) Diagnostic biopsy - wedge/trucut - pipelle/DD&C	Specimens intermediate between 'medium size' & 'very complex'	Radical surgery specimens; especially radical dissection requiring margins and lymph node status, resulting in	a) Complex biopsies:
- Appendix	- skin biopsy(no IF)	Examples:	multiple specimens.	- Lymphoproliferative disorders
- Fallopian tube	- abcess biopsy	- Pneumonectomy/lobectomy		- Liver biopsies (nonneoplastic)
- Vas		- Simple mastectomy; wide	Examples:	- Muscle biopsies
- Tonsils & adenoid	b) Medium size specimens:	local excision/hookwire	- Radical neck dissection	- Renal biopsies (nonneoplastic)
- Sebaceous cyst	- Salivary gland, orchidectomy,	localisation	- Mastectomy with axillary	- Non neoplastic skin biopsies
- Nasal polyps	Lymph node, thyroid, breast	- Gastrectomy; Gut resection.	resection	requiring IF
- Heart valves	lump and omentum for	- Nephrectomy	- Wertheim's hysterectomy	- Biopsies requiring multiple/
- Gallbladder	benign lesions	- Cone biopsy/LLETZ/LEEP	- Vulvectomy with	extensive immunohisto-
- Ganglion	- Eye (lesional excision)	- Thyroid malignancy	lymphadenectomy	chemistry
- POC	- Prostatic chips	- Ovarian malignancy	- Eye exanteration	
- Mucocele	- Splenectomy	- TAHBSO specimen for benign	- Laryngectomy	
 Cervical polyp (benign) 	- Simple hysterectomy/	lesions	- Glossectomy with neck	b) Referral cases:
- Fibroepithelial polyp	myomectomy	- Limb amputation	dissection	- Cases received for second
- Dermoid cyst (skin)	- Simple Ovarian cyst	- Tumour excision >10 cm &/or	- Mandibulectomy/Maxillectomy	opinion
	- Excision of diabetic ulcer	requiring multiple ancillary	- TAHBSO specimen for	
	- Tumour excision < 10 cm	tests	malignant lesions	
	&/or requiring minimum or	- Diagnostic biopsies sent as	- Pelvic exenteration or enbloc	
	no ancillary testing	4-10 separate specimens	resection of multiple organs	
	e.g. neurofibroma, lipoma	- Rectal biopsy for Hirschprung	&/or bowel segments	
	- Diagnostic biopsies as 2-3	Disease		
	separate containers	- Orchidectomy; malignant		
	- Ectopic pregnancy/			
	placenta/ molar pregnancy			
	- Uterus, post partum			

Source: Akta Fi 1951 and Perintah Fi (Perubatan) C (Pesakit Bayar Penuh) 2007

PATHOLOGY SERVICES, MINISTRY OF HEALTH MALAYSIA WORKSHEET FOR ANATOMIC PATHOLOGY WORKLOAD (INDIVIDUAL PATHOLOGIST) 1/2016 – For Laboratory Use Only

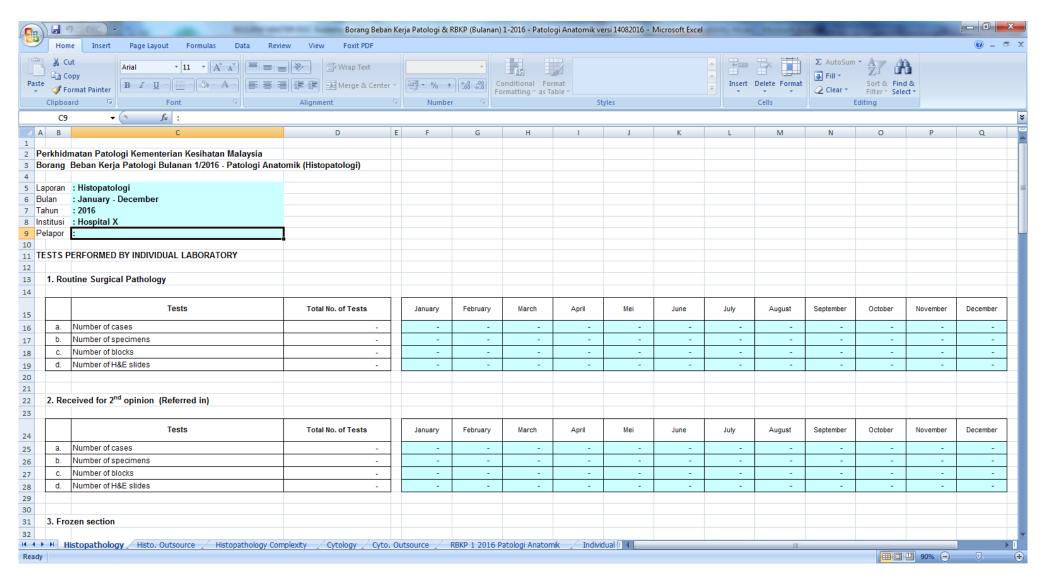
	JAN - JUN	Year		-	
	JAN - DIS	Year:		_	
Hospital:			Reported by:		

No.	NAME	Specify subspecialisation or special posts (e.g.HOD) which may result in less time for daily routine surgical pathology work	*No. of Histopathology Cases Reported & Verified (a)	*No. of Cytopathology Cases Reported & Verified (b)	*Total No. of Histo- & Cyto- Pathology Cases (a+b)	No. of Clinical Autopsy Performed
1						
2						
3						
4						
5						
6						
7						
8						
9						

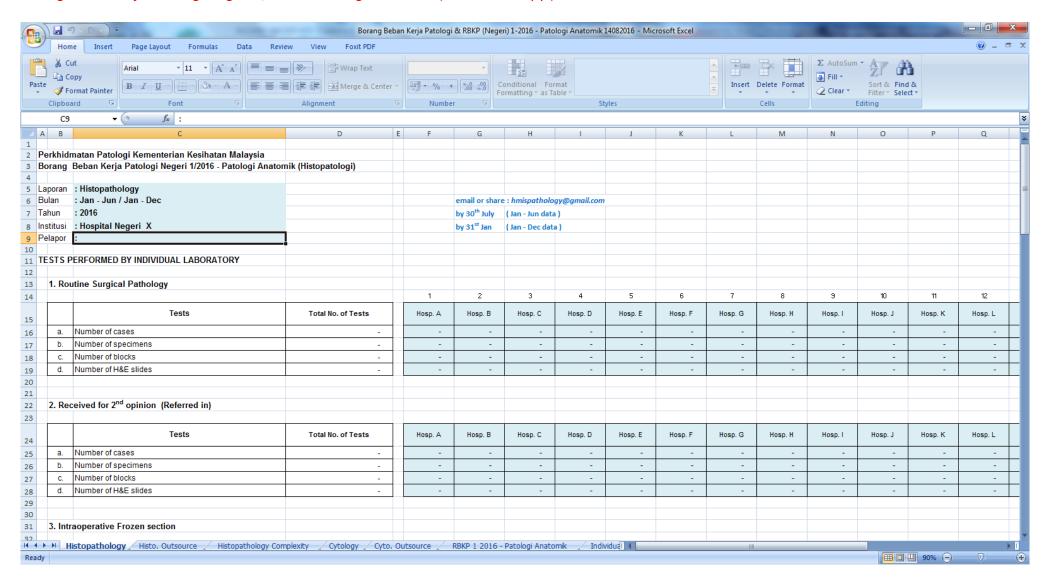
^{*}Note:

Data are only collected for Histopathology and Cytopathology cases that have been reported and verified from 1st until 30th/31st of each month. Cases that are received/registered for the particular month but have yet to be reported are not counted.

Borang Beban Kerja Patologi Bulanan 1/2016 - Patologi Anatomik ('Excel Soft Copy')



Borang Beban Kerja Patologi Negeri 1/2016 - Patologi Anatomik ('Excel Soft Copy')



7.2 CHEMICAL PATHOLOGY

- 7.2.1 In addition to workload reporting to MOH using PER-SS 206 (Pin. 1/2000) form, hospital laboratories are also to submit to their respective State Pathologists in soft copy, detailed, as well as summarised workload data on Chemical Pathology, using Borang Beban Kerja Patologi Bulanan 1/2016 Patologi Kimia and Borang Ringkasan Beban Kerja 1/2016 Patologi Kimia, respectively.
- 7.2.2 The total number of specimens received by a laboratory is calculated from total number of specimens received across the designated groups. These groups are broadly categorised into blood (serum/plasma/whole blood/cord blood/blood spot), urine, body fluid (CSF/sweat/saliva/pleural fluid/peritoneal fluid/pericardial fluid/synovial fluid/vitreous humour etc.) and stool.
- 7.2.3 Each of the test is classified according to the designated sample type and identified accordingly as blood, cord blood, blood spot, urine, CSF, body fluid (peritoneal fluid/pleural fluid/pericardial fluid/synovial fluid etc.), saliva or stool.
- 7.2.4 Each sample run for blood gases (arterial/venous), shall include pH, pCO $_2$ and pO $_2$ as a single test. Other parameters measured by an ABG analyzer are counted separately in the routine chemistry group; e.g. Sodium, Potassium and Ionized Calcium.
- 7.2.5 Tests listed are for automated measurements of analytes, unless stated as non-automated methods e.g. qualitative, semi-quantitative or by dipstick/teststrip.
- 7.2.6 Capillary Bilirubin is the measurement of Total Serum Bilirubin by using spectrophotometer (Bilirubinometer).
- 7.2.7 LDL Cholesterol Direct is considered as one test if it is measured. Calculated LDL Cholesterol is not counted as a test.
- 7.2.8 There are three main categories of routine tests for urine i.e. urine biochemistry, urine for casts and crystals (manual microscopy or automated) and urine for culture and sensitivity (C&S). Only the latter is considered as a truly microbiology test. The final workload for urine biochemistry and urine casts and crystals is to be reported under Chemical Pathology, regardless of where the tests are performed. This principle also applies for urine for eosinophils and urine for dysmorphic rbcs.
- 7.2.9 Urine biochemistry by teststrip/dipstick method may be analysed either manually or using semi-automated or fully automated analyzer. The workload is captured as a single test for each teststrip used, regardless of number of parameters measured and method of analysis.
- 7.2.10 There are several tests generally performed under Chemical Pathology but are also analysed by other disciplines, namely Serology/Immunology. These tests include urine pregnancy test, CRP, C3, C4, IgG, IgA,IgM and IgE. The workload for these tests are captured depending on the local practices and the discipline performing the tests. The final workload however, is to be reported under Chemical Pathology.

- 7.2.11 Each Dynamic Function Test is counted as a single profile and to be considered as one test. The individual test in the dynamic function tests are included/calculated in the respective designated groups.
- 7.2.12 Acylcarnitine profile is included in the blood spot IEM screening.
- 7.2.13 Urine metabolic screening is considered as two tests. The tests include analysis of amino acids and acylcarnitine under positive and negative ion mode which requires different sample preparation and injection into LCMS/MS.
- 7.2.14 For molecular testing, the workload calculation is based on the number of panel of exons per gene tested, i.e. every exon tested in a gene is considered as one test.
- 7.2.15 Drug of Abuse (DOA) screening and confirmatory tests are counted separately as individual tests. For DOA confirmation the parent drug and its metabolites are regarded as individual tests and are reported according to the analytes tested.
- 7.2.16 Any dilution performed for analyte measurement is considered as an additional test and to be included in the workload calculation. The dilution procedure is counted as one additional test, regardless of the number of serial dilutions performed.
- 7.2.17 Any test offered that is not in the list, requires the workload submission to be temporarily added under 'others' and according to designated groups. Please inform National Pathology Workload Committee via Head of Discipline, of the test names that need to be added into the test list in both Borang Beban Kerja Patologi Bulanan 1/2016 Patologi Kimia and Borang Beban Kerja Patologi Negeri 1/2016 Patologi Kimia.
- 7.2.18 Workload calculation and recording:

A. SAMPLE TYPE AND SPECIMEN NUMBER

No.	Sample type	No. of specimens
1	Blood	
2	Urine	
3	Body Fluid	
4	Stool	
	Total number of specimens	

B. ROUTINE CHEMISTRY

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
1	Albumin	Blood		x 1
2	Alkaline Phosphatase (ALP)	Blood		x 1
3	Alkaline Transaminase (ALT)	Blood		x 1
4	Ammonia	Blood		x 1
5	Amylase	Blood		x 1
6	Aspartate Transaminase (AST)	Blood		x 1
7	Bicarbonate	Blood		x 1
8	Bilirubin Direct	Blood		x 1
9	Bilirubin Total	Blood		x 1
10	Bilirubin Total Capillary (Bilirubinometer)	Blood		x 1
11	Calcium Ionised	Blood		x 1
12	Calcium Total	Blood		x 1
13	Chloride	Blood		x 1
14	Cholesterol Total	Blood		x 1
15	Corrected Calcium Total	Blood		x 1
16	Creatine Kinase	Blood		x 1
17	Creatinine	Blood		x 1
18	Gamma Glutamyl Transferase (GGT)	Blood		x 1
	Glucose	Blood		x 1
20	HDL Cholesterol	Blood		x 1
	Ketones	Blood		x 1
	Lactate	Blood		x 1
	Lactate Dehydrogenase (LDH)	Blood		x 1
	LDL Cholesterol Direct (Measured)	Blood	received.	x 1
	Magnesium	Blood		x 1
	Osmolality	Blood		x 1
	Potassium	Blood		x 1
	Phosphate Inorganic	Blood		x 1
	Protein Total	Blood		x 1
	Pyruvate	Blood		x 1
	Sodium	Blood		x 1
	Triglyceride	Blood		x 1
	Urea	Blood		x 1
	Uric Acid (Urate)	Blood		x 1
	Blood Gases (Arterial/Venous)	Blood		x 1
	Albumin	Urine		x 1
	Albumin Creatinine Ratio (UACR)	Urine		x 1
	Amylase/Diastase	Urine		x 1
	Creatinine Clearance	Urine		x 1
	Calcium	Urine		x 1
	Chloride	Urine		x 1
	Creatinine	Urine		x 1
	Fat Globules Urine (Qualitative)	Urine		x 1
	Glucose	Urine		x 1
	Ketones	Urine		x 1
	Magnesium	Urine		x 1
	Osmolality Urine	Urine		x 1
	Potassium	Urine		x 1

49	Protein Creatinine Index (UPCI)	Urine		x 1		
50	Phosphate Inorganic	Urine		x 1		
51	Protein	Urine		x 1		
52	Reducing Sugar Urine	Urine		x 1		
53	Sodium	Urine		x 1		
54	Urea	Urine		x 1		
55	Uric Acid (Urate)	Urine		x 1		
56	Urine Biochemistry (Striptest/Dipstick-Qualitative)	Urine		x 1		
57	Urinary Cast and Crystal	Urine		x 1		
58	Urine for dysmorphic RBC	Urine		x 1		
59	Urine for eosinophil	Urine		x 1		
60	Urine Pregnancy Test (Qualitative)	Urine		x 1		
61	Urine Microalbumin Test Strip (Semi-Quantitative)	Urine		x 1		
62	Urine Microscopy (Manual)	Urine		x 1		
63	Urine Microscopy (Automated)	Urine		x 1		
64	Albumin CSF	CSF		x 1		
65	Chloride CSF	CSF		x 1		
66	Glucose CSF	CSF	Total no. of	x 1		
67	Lactate CSF	CSF	specimens	x 1		
68	Protein CSF	CSF	received.	x 1		
69	Pyruvate CSF	CSF		x 1		
70	Globulin CSF (Qualitative)	CSF		x 1		
71	Albumin Body Fluids	Body Fluids		x 1		
72	Amylase Body Fluids	Body Fluids		x 1		
73	Creatinine Body Fluids	Body Fluids		x 1		
74	Cholesterol Body Fluids	Body Fluids		x 1		
75	Chloride Body Fluids	Body Fluids		x 1		
76	Glucose Body Fluids	Body Fluids		x 1		
77	Urea Peritoneal Dialysate Body Fluids	Body Fluids		x 1		
78	Lactate Dehydrogenase (LDH)	Body Fluids		x 1		
79	pH Body Fluid	Body Fluids		x 1		
80	Potassium Body Fluids	Body Fluids		x 1		
81	Protein, Body Fluids	Body Fluids		x 1		
82	Sodium Body Fluids	Body Fluids		x 1		
83	Fat Globules Stool (Qualitative)	Stool		x 1		
84	Reducing Sugar Stool (Qualitative)	Stool		x 1		
85	Stool occult blood (Qualitative)	Stool		x 1		
86	Others (if test not listed)	Urine		x 1 each		
	TOTAL					

C. ENDOCRINE AND METABOLIC

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
(CI)	ENDOCRINE			
1	17 Hydroxy Progesterone	Blood		x 1
2	Adrenocorticotrophic Hormone (ACTH)	Blood	Total no. of	x 1
3	Aldosterone	Blood	specimens received.	x 1
4	Androstenedione	Blood	70001704.	x 1

	Antidicuratio Harmona (ADH)	Dlaad		
5	Antidiuretic Hormone (ADH)	Blood	-	x 1
6	Anti-Thyroglobulin Antibody	Blood	<u> </u> 	x 1
7	Anti-Thyroid Receptor Antibodies	Blood	-	x 1
8	Beta Cross Laps	Blood	-	x 1
9	Calcitonin	Blood	-	x 1
10	Cortisol	Blood	-	x 1
11	C-Peptide	Blood	 	x 1
12	Dehydroepiandrosterone Sulphate (DHEAS)	Blood	 	x 1
13	Erythropoietin	Blood	<u> </u>	x 1
14	Follicle Stimulating Hormone (FSH)	Blood	-	x 1
15	Gastrin	Blood		x 1
16	Glucagon	Blood		x 1
17	Growth Hormone (Somatotrophin)	Blood		x 1
18	Gonadotrophin Releasing Hormone (GnRH)	Blood		x 1
19	Insulin-like Growth Factor, Binding	Blood		x 1
20	Insulin-like Growth Factor 1 (IGF-1)	Blood		x 1
21	Insulin	Blood		x 1
22	Parathyroid Hormone (intact) 2 nd generation	Blood		x 1
23	Parathyroid Hormone (whole) 3 rd generation	Blood	1	x 1
24	Procalcitonin	Blood		x 1
25	Pro-Insulin	Blood		x 1
26	Luteinising Hormone (LH)	Blood	1	x 1
27	Macroprolactin	Blood	-	x 1
28	Oestradiol	Blood	1	x 1
29	P1NP	Blood	-	x 1
30	Prolactin	Blood	Total no. of	x 1
31	Progesterone	Blood	specimens	x 1
32	Renin	Blood	received.	x 1
33	Sex Hormone Binding Globulins (SHBG)	Blood	-	x 1
34	Tri-lodothyronine (Free T3)	Blood	-	x 1
35	Thyroxine Free (Free T4)	Blood	-	x 1
36	Thyroxine Total (T4)	Blood	-	x 1
37	Testosterone	Blood	-	x 1
38	Thyroglobulin	Blood	-	x 1
39	Thyroid Stimulating Hormone (TSH)	Blood	-	x 1
40	Thyroid Microscomal Antibody	Blood	-	x 1
41	TSH Cord Blood	Cord Blood	-	x 1
42	Free T4 Cord Blood	Cord Blood	-	x 1
43	24-hr Urine 17-OH Keto Steroids	Urine	-	x 1
44	24-hr Urine 17, Ketogenic Steroids	Urine	-	x 1
	24-hr Urine Cathecholamines	Office	1	
	pH Urine		-	x 1
45		-		Captured
	Creatinine Urine - Under Routine Chemistry (RC)	Urine		under RC
	Cathecholamines			x 1
46	24-hr Urine Cortisol	Urine	1	x 1
47	24-hr Urine Free Cortisol	Urine	1	x 1
48	24-hr Urine Pregnanetriol	Urine	1	x 1
	24-hr Urine Metanephrines		1	
	pH Urine		1	x 1
49		I lain -		Captured
	Creatinine Urine - Under Routine Chemistry (RC)	Urine		under RC
	Metanephrines			x 1

50	Midnight Salivary Cortisol	Saliva		x 1
51	Others (if test not listed)			x 1 each
		SUBTOTA	AL ENDOCRINE	
(C II)	METABOLIC			
1	Anti-Glutamic acid decarboxylase (GAD)	Blood		x 1
2	Anti islet cells (ICA)	Blood		x 1
3	Anti-insulin G	Blood		x 1
4	Anti-Insulinoma-Associated Antigen 2 (IA2)	Blood		x 1
5	B 1, Vitamin (Thiamin)	Blood		x 1
6	B 3, Vitamin (Niacin)	Blood		x 1
7	B 6, Vitamin (Pyridoxin)	Blood		x 1
8	B 12, Vitamin	Blood		x 1
9	Beta Carotene	Blood		x 1
10	D, Vitamin	Blood	Total no. of	x 1
11	E, Vitamin	Blood		x 1
12	Ferritin	Blood		x 1
13	Folate	Blood	specimens	x 1
14	Folate RBC	Blood	received.	x 1
15	Fructosamine	Blood		x 1
16	HbA1c (Glycated Hemoglobin)	Blood		x 1
17	Iron, Total	Blood		x 1
18	Iron Binding Capacity, Total (TIBC) - measured	Blood		x 1
19	Iron Binding Capacity, Unsaturated (UIBC) - measured	Blood		x 1
20	Oxalate	Blood		x 1
21	Transferrin	Blood		x 1
22	Iron, Total Urine	Urine		x 1
23	Sweat test	Sweat		x 1
24	Others (if test not listed)			x 1 each
SUBTOTAL METABOLIC				
TOTAL ENDOCRINE AND METABOLIC				

D. CARDIAC MARKERS

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
1	Creatine Kinase Isoenzyme (CKMB) activity	Blood		x 1
2	Creatine Kinase Isoenzyme (CK-MB) Mass	Blood		x 1
3	Troponin-T	Blood	Total no. of	x 1
4	Troponin-I	Blood	specimens received.	x 1
5	Brain Natriuretic Peptide (BNP)	Blood	70007704.	x 1
6	Others (if test not listed)			x 1 each
			TOTAL	

E. TUMOUR MARKERS

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
1	Alpha Feto-Protein (AFP)	Blood	Total no. of specimens received.	x 1
2	Beta Human Chorionic Gonadotrophin	Blood		x 1
3	Cancer Antigen 125 (CA 125)	Blood		x 1
4	Cancer Antigen 15-3 (CA 15-3)	Blood		x 1
5	Cancer Antigen 19-9 (CA 19-9)	Blood		x 1
6	Carcinoembryonic Antigen (CEA)	Blood		x 1
7	Chromogranin A	Blood		x 1
8	Prostate Specific Antigen (PSA) Total	Blood		x 1
9	Prostate Specific Antigen (Free)	Blood		x 1
10	Others (if test not listed)			x 1 each
TOTAL				

F. THERAPEUTIC DRUG MONITORING (TDM)

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
1	Amikacin	Blood		x 1
2	Carbamazepine	Blood		x 1
3	Cyclosporine	Blood		x 1
4	Digoxin	Blood		x 1
5	Everolimus	Blood		x 1
6	Gentamicin	Blood		x 1
7	Lithium	Blood		x 1
8	Methadone Blood	Blood		x 1
9	Methaqualone	Blood	Total no. of	x 1
10	Methotrexate (MTX)	Blood		x 1
11	Netilmicin	Blood	specimens received.	x 1
12	Phenobarbital	Blood		x 1
13	Phenytoin (Dilantin)	Blood		x 1
14	Sirolimus	Blood		x 1
15	Tacrolimus	Blood		x 1
16	Theophylline	Blood		x 1
17	Valproic acid	Blood		x 1
18	Vancomycin	Blood		x 1
19	Methadone Urine	Urine		x 1
20	Others (if test not listed)			x 1 each
TOTAL				

G. TOXICOLOGY

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
1	Acetaminophen	Blood	Total no. of specimens received.	x 1
2	Alcohol (Ethanol)	Blood		x 1
3	Benzodiazepine	Blood		x 1
4	Cholinesterase	Blood		x 1
5	Methanol	Blood		x 1

6	Salicylate	Blood		x 1	
7	Alcohol (Ethanol) Urine	Urine	Total no. of specimens received.	x 1	
8	Paraquat Urine	Urine		x 1	
9	Salicylate Urine	Urine		x 1	
10	Others (if test not listed)			x 1 each	
TOTAL					

H. DRUG OF ABUSE (DOA)

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
1	Adulteration test; Creatinine	Urine	-	x 1
2	Adulteration test; Specific Gravity	Urine		x 1
3	Adulteration test; pH	Urine		x 1
4	Amphetamine screening	Urine		x 1
5	Amphetamine confirmation	Urine		x 1
6	Benzodiazepines screening	Urine		x 1
7	Benzoylecgonine	Urine		x 1
8	Cannabinoids screening	Urine		x 1
9	Cannabinoids confirmation	Urine		x 1
10	Cocaine screening	Urine		x 1
11	Cocaine confirmation	Urine		x 1
12	Codeine	Urine		x 1
13	Dextromethorphan	Urine		x 1
14	Ephedrine	Urine		x 1
15	Hydroxy-norketamine	Urine		x 1
16	Ketamine screening	Urine		x 1
17	Ketamine confirmation	Urine	Total no. of	x 1
18	Lysergic acid diethylamide (LSD) screening	Urine		x 1
19	Lysergic acid diethylamide (LSD) confirmation	Urine	specimens received.	x 1
20	Methamphetamine screening	Urine	received.	x 1
21	Methamphetamine confirmation	Urine		x 1
22	Methylenedioxyethylamphetamine (MDEA)	Urine		x 1
23	Methylenedioxymethamphetamine (MDMA)	Urine		x 1
24	Monoacetylmorphine screening	Urine		x 1
25	Monoacetylmorphine confirmation	Urine		x 1
26	Morphine confirmation	Urine		x 1
27	N-methyl-1,3-benzodioxolylbutanamine (MBDB)	Urine		x 1
28	Norephedrine	Urine		x 1
29	Norketamine	Urine		x 1
30	Nimetazepam	Urine		x 1
31	Nitrazepam	Urine		x 1
32	Opiate screening	Urine		x 1
33	Phencyclidine (PCP) screening	Urine		x 1
34	Phencyclidine (PCP) confirmation	Urine		x 1
35	Phentermine	Urine		x 1
36	Others (if test not listed)			x 1 each
			TOTAL	

I. DYNAMIC FUNCTION TEST

No.	Dynamic Function Test (DFT)	No. of DFT
1	ACTH stimulation for Congenital Adrenal Hyperplasia (CAH)	1 DFT
2	Adrenal Venous Sampling (AVS)	1 DFT
3	Arterial Stimulation and Venous Sampling (ASVS)	1 DFT
4	Aldosterone Renin Ratio (ARR)	1 DFT
5	Ammonium Chloride Loading Test	1 DFT
6	Captopril Challenge Test	1 DFT
7	Combined Anterior Pituitary Function Test	1 DFT
8	CRH Stimulation Test	1 DFT
9	Fludrocortisone Suppression Test	1 DFT
10	Free Androgen Index (FAI)	1 DFT
11	Gonadotropin Releasing hormone Stimulation Test	1 DFT
12	Glucagon Stimulation Test	1 DFT
13	High Dose Dexamethasone Suppression Test (HDDST)	1 DFT
14	Human Chorionic Gonadotrophin Test (hCG test)	1 DFT
15	Inferior Petrosal Sinus Sampling (IPSS)	1 DFT
16	Insulin Tolerance Test	1 DFT
17	Low Dose Dexamethasone Suppression Test	1 DFT
18	Metoclopramide Stimulation Test	1 DFT
19	Overnight Dexamethasone Suppression Test (ODST)	1 DFT
20	Saline Suppression Test	1 DFT
21	Short Synacthen Test	1 DFT
22	Water Deprivation Test	1 DFT
23	Water Loading Test	1 DFT
24	Others (if test not listed)	1 DFT each
	TOTAL	

J. SPECIAL PROTEIN AND PROTEOMICS

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
1	Alpha-1-Antitrypsin-Quantitation	Blood		x 1
	Alpha-1-Antitrypsin-Phenotyping			
2	Alpha-1-Antitrypsin-Quantitation	Blood	Total no. of specimens received.	x 1
	Alpha-1-Antitrypsin-Electrophoresis gel	DIOOG		x 1
3	Alpha-1-Acid Glycoprotein (Orosomucoids)	Blood		x 1
4	Alpha 2 Macroglobulin	Blood		x 1
5	Apolipoprotein A-1	Blood		x 1
6	Apolipoprotein B	Blood		x 1
7	Apolipoprotein C	Blood		x 1
8	Apolipoprotein D	Blood		x 1
9	Apolipoprotein E	Blood		x 1
10	Apolipoprotein E (Phenotyping)	Blood		x 1
11	Beta-2 Microglobulin	Blood		x 1
12	Complement 3 (C3)	Blood		x 1

13	Complement 4 (C4)	Blood		x 1	
14	Caeruloplasmin	Blood		x 1	
15	C-Reactive Protein (CRP)	Blood		x 1	
16	Cryoglobulin screening	Blood		x 1	
10	Cryoglobulin	Blood			x 1
17	Plasma Electrophoresis and Immunofixation	Blood		Captured under	
''	Serum Electrophoresis and Immunofixation	Blood	-	SPE and IF	
18	Free Kappa Light Chain Serum	Blood	1	x 1	
19	Free Lambda Light Chain Serum	Blood	1	x 1	
20	Haptoglobin	Blood	1	x 1	
21	Immunoglobulin A (IgA)	Blood	1	x 1	
22	Immunoglobulin E (IgE)	Blood	1	x 1	
23	Immunoglobulin G (IgG)	Blood	1	x 1	
24	Immunoglobulin M (IgM)	Blood	1		
25	Lipoprotein (a) Electrophoresis	Blood	-	x 1 x 1	
26	Myoglobin Serum	Blood	_	x 1	
	, ,	Blood	-		
27	Pre Albumin Quantitative		-	x 1	
28	Transferrin Isoform	Blood	_	x 1	
00					
29	Total Protein	Blood		x 1	
	Electrophoresis Gel			x 1	
20	Protein Electrophoresis Serum		Total no. of specimens	1	
30	Total Protein Serum	Blood		x 1	
	Protein Electrophoresis Gel Test Protein Immunofixation Serum			x 1	
			received.	v 1	
	Albumin			x 1 x 1	
	Immunoglobulin G (IgG)	Dlood			
31	Immunoglobulin A (IgA)	Blood		x 1	
	Immunoglobulin M (IgM)			x 1	
	Kappa Lambda			x 1	
	Immunofixation Gel Test			x 1	
				x 1	
32	Protein Immunofixation Serum (D,E) Immunofixation Gel Test		-		
32	(5 antisera: IgG,IgD,IgE, Kappa & Lambda)			x 1	
33	Free Kappa Light Chain Urine	Urine		x 1	
34	Free Lambda Light Chain Urine	Urine		x 1	
<u> </u>	Protein Electrophoresis Urine	00		X /	
	Concentrate Urine			x 1	
35	Total Protein Urine	Urine		x 1	
	Protein Electrophoresis Gel Test			x 1	
	Protein Immunofixation Urine			X /	
	Albumin			x 1	
	Immunoglobulin G (IgG)	Urine		x 1	
	Immunoglobulin A (IgA)			x 1	
36	Immunoglobulin M (IgM)			x 1	
	Kappa	Oillie		x 1	
	Lambda			x 1	
	Immunofixation Gel Test	_		x 1	
	i iiiiiidiidiixalidii Gel Test			^ /	

	Beta-2 Microglobulin Urine			
37	Beta-2 Microglobulin Urine	Urine		x 1
31	Beta-2 Microglobulin Serum	Blood		x 1
	Myoglobin Urine	Urine		x 1
	Protein Electrophoresis CSF			
	Albumin CSF	CSF		x 1
	IgG CSF		Tatalina of	x 1
38	Total Protein CSF		Total no. of specimens - received	x 1
30	CSF Protein Electrophoresis Gel (CSF & Serum)			x 1
	Albumin Serum	Blood		x 1
	IgG Serum			x 1
	Total Protein Serum			x 1
	Beta-2 Microglobulin CSF			
39	Beta-2 Microglobulin CSF	CSF		x 1
	Beta-2 Microglobulin Serum	Blood		x 1
40	Lecithin/Sphingomyelin Ratio Amniotic Fluid	Body Fluid		x 1
41	Others (if test not listed)			x 1 each
			TOTAL	

K. ENZYMOLOGY

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
1	Acid Phosphatase	Blood		x 1
2	Alkaline Phosphatase Isoenzymes	Blood		x 1
3	Alkaline Phosphatase Bone Spesific	Blood	Total no. of	x 1
4	Aldolase	Blood	specimens	x 1
5	Erythrocyte transketolase	Blood	received.	x 1
6	Lactate Dehydrogenase Iso-enzymes	Blood		x 1
7	Others (if test not listed)			x 1
			TOTAL	

L. TRACE ELEMENTS

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
1	Aluminium	Blood		x 1
2	Cadmium	Blood		x 1
3	Chromium	Blood		x 1
4	Copper	Blood		x 1
5	Fluoride	Blood		x 1
6	Lead	Blood	Total no of	x 1
7	Mercury	Blood	specimens received.	x 1
8	Selenium	Blood	70007704.	x 1
9	Zinc	Blood		x 1
10	Aluminium Urine	Urine		x 1
11	Arsenic Urine, 24H	Urine		x 1
12	Benzoylecgonine Urine	Urine		x 1

13	Cadmium Urine	Urine		x 1
14	Copper Urine, 24H	Urine		x 1
15	Fluoride Urine	Urine	1	x 1
16	Iodine Urine	Urine	1	x 1
17	Lead Urine, 24H	Urine	1	x 1
18	Mercury Urine	Urine	1	x 1
19	Aluminium	Body Fluid		x 1
20	Antimony	Body Fluid		x 1
21	Arsenic	Body Fluid		x 1
22	Barium	Body Fluid		x 1
23	Beryllium	Body Fluid		x 1
24	Cadmium	Body Fluid	Total no of	x 1
25	Chloramines	Body Fluid	specimens	x 1
26	Chromium	Body Fluid	received.	x 1
27	Copper	Body Fluid	Toocivea.	x 1
28	Fluoride	Body Fluid		x 1
29	Free Chlorine	Body Fluid		x 1
30	Lead	Body Fluid		x 1
31	Mercury	Body Fluid		x 1
32	Selenium	Body Fluid		x 1
33	Silver	Body Fluid		x 1
34	Sulphate	Body Fluid		x 1
35	Thallium	Body Fluid	[x 1
36	Zinc	Body Fluid		x 1
37	Lead Hair	Hair		x 1
38	Others (if test not listed)			x 1 each
			TOTAL	

M. BIOCHEMICAL GENETICS

No.	Test Name	Sample Type	No. Specimens	No. of Test performed
1	Acid alpha-Glucosidase	Blood Spot		x 1
2	Galactose 1-Uridyl transferase	Blood Spot		x 1
3	Galactose, Total	Blood Spot		x 1
4	IEM screening (Blood Spot)	Blood Spot		x 1
5	Lysosomal Storage disease (LSD) screening	Blood Spot		x 1
6	Biotinidase enzyme activity	Blood Spot		x 1
7	Amino acid Plasma	Blood		x 1
8	Carnitine, Free Plasma	Blood		x 1
9	Carnitine, Total Plasma	Blood	Total no. of	x 1
10	CDG phenotyping	Blood	specimens	x 1
11	α-fucosidase	Blood	received.	x 1
12	α-galactosidase	Blood		x 1
13	α-hexosaminidase (AHEX	Blood		x 1
14	α-iduronidase (IDA)	Blood		x 1
15	α-mannosidase (AMAN)	Blood		x 1
16	α-N-acetyl-galactosaminidase	Blood		x 1
17	Aryl Sulphatase A	Blood		x 1
18	Aspartyl Glucosaminidase (GASP)	Blood		x 1
19	Branching enzyme (GSD IV)	Blood		x 1

67	Others (if test not listed)			x 1 each
		i e		4
66	Urobilinogen	Urine		x 1
65	5-hydroxy-Indol-Acetic Acid (5 HIAA) 24Hr Urine	Urine		x 1
64	Sulphite	Urine		x 1
63	Sulfocysteine	Urine		x 1
62	Sugars and polyols	Urine		x 1
61	Succinylacetone	Urine		x 1
60	Sialic acids Free Urine	Urine		x 1
59	Sialic acids Total Urine	Urine		x 1
58	Purine & Pyrimidine Urine	Urine		x 1
57	Pterins Urine	Urine		x 1
55 56	Porphobilinogen Urine Porphyrins Urine	Urine Urine		x 1 x 1
54	Pipecolic Parabability and I Itria	Urine		x 1
53	Orotic Acid (Orotate) Urine	Urine		x 1
52	Organic Acids Urine	Urine		x 1
51	Oligosaccharides Urine	Urine		x 1
50	ResolutionElectrophoresis) Urine	Urine		x 1
	Mucopolysacharides (High			
49	Methylmalonic acid	Urine		x 1
	Acylcarnitine	Urine		x 1
48	Amino Acids	Urine		x 1
	IEM Screening, Urine			
47	Homocystine Urine (Qualitative)	Urine		x 1
46	Glycosaminoglycans (Quantitation)	Urine		x 1
45	Delta Amino Laevulinic Acid Urine, 24Hr Urine	Urine		x 1
44	Cystine Urine (Qualitative)	Urine	received.	x 1
43	Creatine	Urine	specimens	x 1
42	Carnitine, 24Hr Urine	Urine	Total no. of	x 1
41	Biogenic Amines Urine	Urine		x 1
40	Amino acid Urine	Urine		x 1
39	Alpha-aminoadipic semialdehyde	Urine		x 1
38	Pterins CSF	CSF		x 1
37	Biogenic Amines CSF	CSF		x 1
36	Amino acid CSF	CSF		x 1
35	VLCFA & Phytanic acid Plasma	Blood		x 1
34	Pipecolic acid Plasma	Blood		x 1
33	Homocysteine Total, Plasma	Blood		x 1
32	Palmitoyl Protein Thioesterase	Blood		x 1
	Arylsuphatase B (ASB)			
31	N-acetylgalactosamine 4-sulphatase /	Blood		x 1
30	N-acetylgalactosamine-6-sulfatase (GALSO)	Blood		x 1
29	Methylmalonic acid	Blood		x 1
28	Iduronate-2-sulphatase (IDS)	Blood		x 1
27	Heparan Sulphamidase (SULP)	Blood		x 1
26	Galactocerebrosidase (GALC)	Blood		x 1
25	β-mannosidase (BMAN)	Blood		x 1
24	β-hexosaminidase (BHEX)	Blood		x 1
23	β-glucosidase	Blood		x 1
22	β-galactosidase	Blood		x 1
21	β-hexosaminidase A (MUGS)	Blood		x 1
20	β-glucuronidase (BGLUCU)	Blood		x 1

N. MOLECULAR GENETICS

Method Classification:

M : Manual

S : Semi automation F : Full automation

No.	Test Name	Sample Type	No. Specimens	Method M/S/F	No. of Test performed
1	Acute Intermittent Porphyria (HMBS) Sequencing	Blood			x No. of test panel tested
2	Acute Intermittent Porphyria (HMBS) MLPA	Blood	1		x No. of test
		5.000	-		panel tested
3	Alagille Syndrome (JAG1) Sequencing	Blood			x No. of test panel tested
4	Alagille Syndrome (JAG1) MLPA	Blood	1		x No. of test panel tested
5	Alagille Syndrome (NOTCH2)	Blood	† †		x No. of test
6		Blood	-		panel tested x No. of test
0	Alexander Disease (GFAP)	Біооц	4		panel tested
7	Alpha 1-Antitrypsin Deficiency (SERPINA1)	Blood			x No. of test panel tested
8	Angelman Syndrome (SNRPN) MS-PCR	Blood			x No. of test
			-		panel tested x No. of test
9	Angelman Syndrome (UBE3A) Sequencing	Blood			panel tested
10	Angelman Syndrome (UBE3A) MLPA	Blood			x No. of test panel tested
11	Argininosuccinate Lyase Deficiency (ASL)	Blood	-		x No. of test
11		ыооч	_		panel tested
12	Argininosuccinate Synthase Deficiency (ASS1)	Blood			x No. of test panel tested
13	Aromatic Amino Acid Decarboxylase Deficiency (DDC)	Blood	Total no. of		x No. of test panel tested
14	Berardinelli Congenital Lipodystrophy (BSCL2)	Blood	specimens received.		x No. of test panel tested
15	Berardinelli Congenital Lipodystrophy (AGPAT2)	Blood			x No. of test panel tested
16	Biotinidase Deficiency (<i>BTD</i>)	Blood			x No. of test panel tested
17	BRCA-1 & 2	Blood			x No. of test panel tested
18	CADASIL (NOTCH3) - hotspots	Blood			x No. of test panel tested
19	Canavan Disease (ASPA)	Blood	1		x No. of test
	Carbamoylphosphate Synthetase 1		-		panel tested x No. of test
20	Deficiency (CPS1)	Blood			panel tested
21	Carnitine Update Deficiency (OCTN2)	Blood			x No. of test panel tested
22	Carnitine-Acylcarnitine Translocase Deficiency (<i>SLC25A20</i>)	Blood	1		x No. of test panel tested
23	Carnitine Palmitoyltransferase 1A (CPT1) Deficiency (CPT1A)	Blood			x No. of test panel tested
24	Carnitine Palmitoyltransferase II (CPT 2) Deficiency (CPT2)	Blood			x No. of test panel tested
25	Citrin Deficiency (SLC25A13)	Blood	† †		x No. of test panel tested
26	Classical Homocystinuria (CBS)	Blood	† †		x No. of test
-					panel tested

27	Dihydropyrimidinase (DHP) Deficiency (DPYS)	Blood		x No. of test panel tested
28	DNA Extraction & Storage	Blood		x No. of test panel tested
29	Ethylmalonic Encephalopathy (ETHE1)	Blood	-	x No. of test
			<u> </u>	panel tested x No. of test
30	Fragile X Syndrome (FRAXA) (FMR1)	Blood		panel tested
31	Fragile X Syndrome (FRAXE) (FMR2)	Blood		x No. of test panel tested
32	Fructose-1,6-Bisphosphatase Deficiency (FBP1)	Blood		x No. of test panel tested
33	Fucosidosis (FUCA1)	Blood		x No. of test panel tested
34	Floating-Harbor Syndrome (FHS) (SRCAP)	Blood		x No. of test panel tested
35	Galactokinase Deficiency (GALK1)	Blood		x No. of test panel tested
36	Galactose Epimerase Deficiency (GALE)	Blood		x No. of test panel tested
37	Gaucher Disease (GBA)	Blood		x No. of test panel tested
38	Glutaric Aciduria Type 1 (GCDH)	Blood		x No. of test panel tested
39	Glycogen Storage Disease Type I (GSDI)	Blood		x No. of test
40	(G6P6) Glycogen Storage Disease Type I (GSDI)	Blood		panel tested x No. of test
	(SLC37A4) Glycogen Storage Disease Type III (GSDIII)	2.000	-	panel tested x No. of test
41	(AGL)	Blood	Total no. of	panel tested
42	Hereditary Orotic Aciduria (UMPS)	Blood	specimens	x No. of test panel tested
43	Hypophosphatasia (ALPL)	Blood	received.	x No. of test panel tested
44	Isolated Methyl Malonic Aciduria (MMA) (MUT)	Blood		x No. of test panel tested
45	Isolated Methyl Malonic Aciduria (MMA) (MMAA)	Blood		x No. of test panel tested
46	Isolated Methyl Malonic Aciduria (MMA) (MMAB)	Blood		x No. of test panel tested
47	Leber's hereditary optic neuropathy (LHON)	Blood		x No. of test panel tested
48	Leigh Syndrome (SURF1)	Blood		x No. of test panel tested
49	Leigh Syndrome (8993 hotspot)	Blood	1	x No. of test panel tested
50	Leigh Syndrome (mtDNA Full panel)	Blood		x No. of test panel tested
51	Leopard Syndrome (PTPN11)	Blood		x No. of test panel tested
52	Lesch-Nyhan Syndrome (HPRT)	Blood		x No. of test panel tested
53	Lissencephaly (LIS1)	Blood		x No. of test panel tested
54	Lissencephaly (DCX)	Blood		x No. of test panel tested
55	Long-Chain 3-Hydroxyacyl-CoA	Blood		x No. of test
56	Dehydrogenase (<i>HADHA</i>) Lysinuric Protein Intolerance (<i>SLC7A7</i>)	Blood	-	panel tested x No. of test
			-	panel tested x No. of test
57	Maple Syrup Urine Disease (BCKDHA)	Blood		panel tested

58	Maple Syrup Urine Disease (BCKDHB)		-	x No. of test panel tested
59	Maple Syrup Urine Disease (<i>DBT</i>)	Blood		x No. of test panel tested
60	Maple Syrup Urine Disease (DLD)	Blood		x No. of test
	Maroteaux-Lamy Syndrome, MPS VI		1	panel tested x No. of test
61	(ARSB) MCT8-Specific Thyroid Hormone Cell	Blood	_	panel tested
62	Transporter Deficiency (SLC16A2)	Blood		x No. of test panel tested
63	Medium Chain Acyl-CoA Dehydrogenase (MCAD) Deficiency (<i>ACADM</i>)	Blood		x No. of test panel tested
64	Metachromatic Leukodystrophy (MLD) (ARSA)	Blood		x No. of test panel tested
65	Methylenetetrahydrofolate Reductase Deficiency (<i>MTHFR</i>)	Blood		x No. of test panel tested
66	Methylmalonic Aciduria and Homocystinuria, cbIC Type (MMACHC)	Blood		x No. of test panel tested
67	Methylmalonic Aciduria and Homocystinuria Type D (<i>MMADHC</i>)	Blood		x No. of test panel tested
68	Methylmalonyl-CoA Epimerase Deficiency (MCEE)	Blood		x No. of test panel tested
69	mtDNA Deletion Syndromes - Kearns-Sayre Syndrome (KSS) MLPA	Blood		x No. of test panel tested
70	mtDNA Deletion Syndromes - Pearson Syndrome MLPA	Blood		x No. of test panel tested
71	mtDNA Deletion Syndromes – Chronic Progressive External Ophthalmoplegia (CPEO) MLPA	Blood	Total no. of	x No. of test panel tested
72	mtDNA Depletion Syndrome (MDS) Panel - ANT1	Blood	specimens received.	x No. of test panel tested
73	mtDNA Depletion Syndrome (MDS) Panel - DGUOK	Blood	_ received.	x No. of test panel tested
74	mtDNA Depletion Syndrome (MDS) Panel - MPV17	Blood		x No. of test panel tested
75	mtDNA Depletion Syndrome (MDS) Panel - POLG	Blood		x No. of test panel tested
76	mtDNA Depletion Syndrome (MDS) Panel - RRM2B	Blood		x No. of test panel tested
77	mtDNA Depletion Syndrome (MDS) Panel – SUCLA2	Blood	_	x No. of test panel tested
78	mtDNA Depletion Syndrome (MDS) Panel – SUCLG1	Blood		x No. of test panel tested
79	mtDNA Depletion Syndrome (MDS) Panel – TWINKLE	Blood		x No. of test panel tested
80	mtDNA Depletion Syndrome (MDS) Panel – TYMP	Blood		x No. of test panel tested
81	Mitochondrial Encephalomyopathy, Lactic Acidosis, and Stroke-Like Episodes (MELAS) Syndrome (3243 hotspot)	Blood		x No. of test panel tested
82	Mitochondrial Encephalomyopathy, Lactic Acidosis, and Stroke-Like Episodes (MELAS) Syndrome (full panel)	Blood		x No. of test panel tested
83	Mitochondrial Short-Chain Enoyl-CoA Hydratase 1 Deficiency (ECHS1)	Blood		x No. of test panel tested
84	Morquio A Disease (MPS IVA) - GALNS	Blood		x No. of test panel tested
85	Multiple Respiratory Chain Deficiencies (Mitochondrial Translation Defect) (<i>GFM1</i>)	Blood		x No. of test panel tested

86	Myoclonic Epilepsy with Ragged-Red Fibers (MERRF) Syndrome	Blood		x No. of test panel tested
87	N-Acetylglutamate Synthase (NAGS) Deficiency (NAGS)	Blood		x No. of test panel tested
88	Neuropathy, Ataxia and Retinitis Pigmentosa (<i>NARP</i>) Syndrome	Blood		x No. of test panel tested
89	Non Ketotic Hyperglycinemia (NKH) (<i>AMT</i>)	Blood		x No. of test panel tested
90	Non Ketotic Hyperglycinemia (NKH) (GCSH)	Blood	_	x No. of test panel tested
91	Non Ketotic Hyperglycinemia (NKH) (GLDC)	Blood		x No. of test
92	Non Ketotic Hyperglycinemia (NKH) (GLDC)	Blood	_	panel tested x No. of test
93	MLPA Noonan Syndrome (<i>PTPN11</i>)	Blood		panel tested x No. of test
94	Ornithine Transcarbamylase (OTC)	Blood		panel tested x No. of test
	Deficiency (OTC) Phosphomannomutase 2 Deficiency			panel tested x No. of test
95	(PMM2-CDG) (PMM2)	Blood		panel tested
96	POLG-Related Disorders	Blood		x No. of test panel tested
97	Pompe Disease (GAA)	Blood		x No. of test panel tested
98	Prader-Willi Syndrome (SNRPN) MS-PCR	Blood		x No. of test panel tested
99	Primary Dystonia - THAP1 (DYT6)	Blood		x No. of test panel tested
100	Primary Dystonia – TOR1A (DYT1)	Blood	Total no. of	x No. of test panel tested
101	Pseudorheumatoid Dysplasia (WISP3)	Blood	specimens received.	x No. of test panel tested
102	PTEN-associated Diseases (PTEN) Sequencing	Blood		x No. of test panel tested
103	PTEN-associated Diseases (<i>PTEN</i>) MLPA	Blood		x No. of test panel tested
104	Purine Nucleoside Phosphorylase Deficiency (<i>PNP</i>)	Blood		x No. of test panel tested
105	Pyruvate Dehydrogenase Deficiency (PDHA1)	Blood		x No. of test panel tested
106	Retinoblastoma (RB1) Sequencing	Blood		x No. of test panel tested
107	Retinoblastoma (RB1) MLPA	Blood		x No. of test panel tested
108	Schinzel Giedion Syndrome (SETBP1)	Blood		x No. of test panel tested
109	SCN1A-Related Seizure Disorders (SCN1A)	Blood	1	x No. of test panel tested
110	Severe Congenital Neutropenia (<i>ELANE</i>)	Blood		x No. of test panel tested
111	Short-Chain 3-Hydroxyacyl-CoA Dehydrogenase (SCHAD) Deficiency (HADH)	Blood		x No. of test panel tested
112	Spinal Muscular Atrophy (SMA) Sequencing	Blood		x No. of test panel tested
113	Spinal Muscular Atrophy (SMA) MLPA	Blood		x No. of test panel tested
114	Spinal Muscular Atrophy (SMA) PCR-RFLP	Blood	1	x No. of test panel tested
115	Sulfite Oxidase (SUOX) Deficiency (SUOX)	Blood		x No. of test panel tested
L		<u> </u>	1	paner testeu

116	Tyrosine Hydroxylase Deficiency (<i>TH</i>)	Blood		x No. of test panel tested
117	Very Long Chain Acyl-CoA Dehydrogenase (VLCAD) Deficiency (ACADVL)	Blood		x No. of test panel tested
118	Whole mitochondrial DNA (Full panel)	Blood		x No. of test panel tested
119	Whole mitochondrial DNA (mtDNA hotspots)	Blood		x No. of test panel tested
120	X-Chromosome Inactivation	Blood	Total no. of specimens	x No. of test panel tested
121	X-linked Adrenoleukodystrophy (ABCD1)	Blood	received.	x No. of test panel tested
122	Spinal Muscular Atrophy (SMN) Nuclear Gene Sequence	Blood/blood spot		x No. of test panel tested
123	Duchenne Muscular Dystrophy	Blood		x No. of test panel tested
124	Others (if test not listed)			x No. of test panel tested for each test
TOTAL				

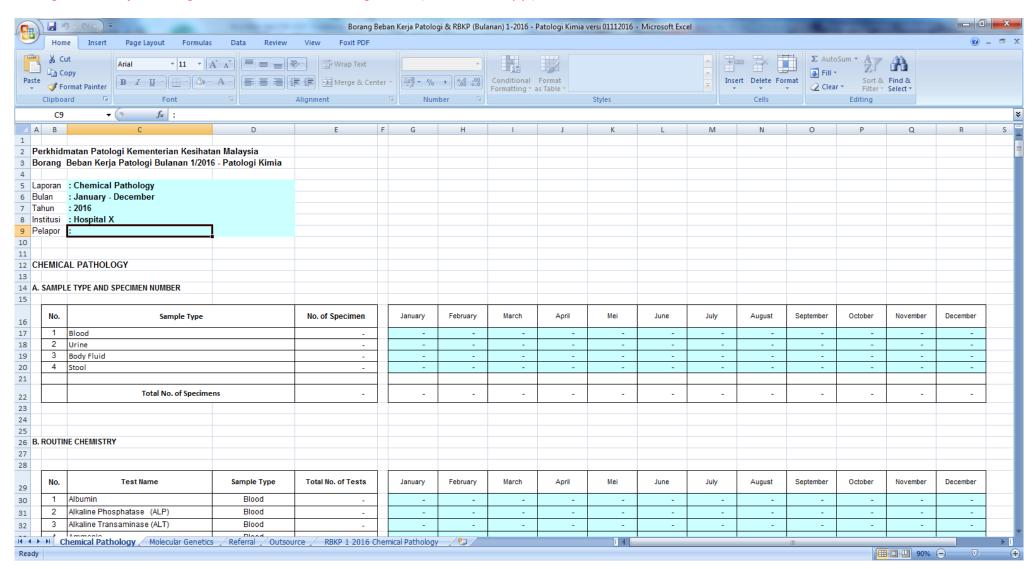
PATHOLOGY SERVICES MINISTRY OF HEALTH, MALAYSIA

REPORT ON LABORATORY WORKLOAD: CHEMICAL PATHOLOGY

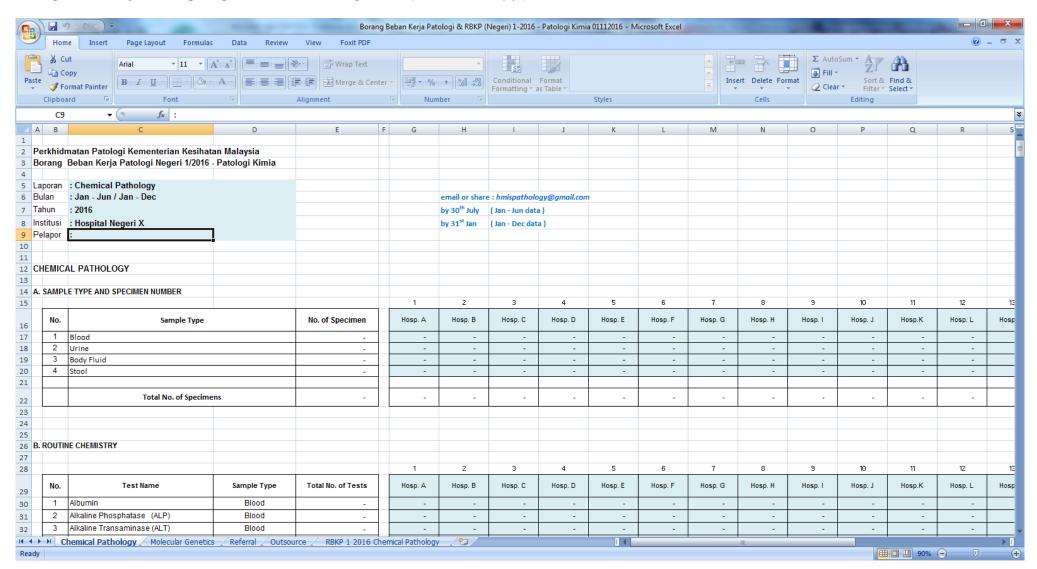
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Month or Location of Hospitals, Institutions and Public Health Facilities	Routine Chemistry	Endocroine & Metabolic	Cardiac Markers	Tumour Markers	Therapeutic Drug Monitoring (TDM)	Toxicology	Drug of Abuse (DOA)	Dynamic Function Test	Protein & Proteomic	Enzymology	Trace Elements	Biochemical Genetics	Molecular Genetics	Total No. of Specimens	Total No. of Tests

Borang Beban Kerja Patologi Bulanan 1/2016 – Patologi Kimia ('Excel Soft Copy')



Borang Beban Kerja Patologi Negeri 1/2016 – Patologi Kimia ('Excel Soft Copy')



7.3 HAEMATOLOGY

- 7.3.1 In addition to workload reporting to MOH using PER-SS 206 (Pin. 1/2000) form, hospital laboratories are also to submit to their respective State Pathologists in soft copy, detailed, as well as, summarised workload data on Haematology, using Borang Beban Kerja Patologi Bulanan 1/2016 Hematologi and Borang Ringkasan Beban Kerja Patologi 1/2016 Hematologi, respectively.
- 7.3.2 The total number of specimens received by a laboratory is calculated from the total number of specimens received across the designated groups.
- 7.3.3 Where more than one specimen are received for one particular test request, the number of specimens to be captured in the workload will depend on testing performed on these specimens; e.g. when peripheral blood and bone marrow aspirate (BMA) are received in 2 separate EDTA tubes from one patient for leukaemia/lymphoma immunophenotyping but the screening and confirmatory tests are performed on BMA only, the number of specimens received for this case is counted as one. The specimen that is not used for testing is disregarded.
- 7.3.4 Blood counts in profiles (e.g DIVC screening and acute leukaemia screening by immunophenotyping) and reticulocyte count in profiles (e.g Hb Analysis is) are counted under FBC and reticulocyte count in routine haematology. In almost all laboratories, the blood counts and automated reticulocyte counts in the profile tests are performed on the same routine FBC haematology analyzer. This will avoid duplication of data and helps to capture the true FBC workload.
- 7.3.5 Reticulocyte count is classified according to 2 methods i.e. automated or manual but similarly counted as one test.
- 7.3.6 Each profile test in general haematology and basic haemostasis and thrombosis is counted as a single profile test. However, the tests in these profiles (indicated by grey-coloured cells below) are individually counted within the designated group. Examples are blood and reticulocyte counts in FBP, and PT and APTT in DIVC screening. This will reflect the true workload of basic haematology services, which consist mainly of blood counts and are available in all laboratories in MOH hospitals and health clinics. Basic coagulation tests PT/INR and APTT are provided in all MOH hospitals.
- 7.3.7 Calculation of workload for profile tests in specialised haematology i.e. other than general haematology and basic haemostasis and thrombosis, is based on the individual tests in each profile. Any additional test is individually counted as one test in the profile e.g. for Hb Analysis, gel electrophoresis in alkaline phase is one test and in acid phase is one additional test. Every monoclonal antibody used in immunophenotyping is considered as one test.
- 7.3.8 Bone marrow aspiration as a single profile is counted as one test. Other than the routine stains, each additional cytochemical stain performed is individually counted as one test (including myeloperoxidase stain performed in leukaemia/lymphoma

- immunophenotyping). Control slides (slides separately stained from test samples) are not counted to ensure more accurate workload data collection.
- 7.3.9 Processing of bone marrow trephine biopsy is performed in Histopathology lab. Their reports are however, included under Haematology specialists' reporting workload, if reported by Haematologists.
- 7.3.10 Workload calculation for ABO and Rhesus blood grouping (immunohaematology) is captured for primary healthcare services (*Klinik Kesihatan*) only.
- 7.3.11 The workload for cerebrospinal fluid (CSF) cytology testing (including cytospin) is captured depending on the local practices and the specialists reporting the test. In many laboratories, CSF cytospin for blasts is offered, performed and reported by haematopathologists, whether as an individual test or as part of screening in immunophenotyping in leukaemia/lymphoma. CSF cytology other than for blasts is under Cytology services.
- 7.3.12 The workload for molecular testing, is classified according to test method, i.e. manual, semi-automation or full automation.
- 7.3.13 For quantitative molecular testing in which, duplication of tests is mandatory for every primer, each primer used is countered as one test, regardless of the number of repeats.
- 7.3.14 Workload calculation for molecular tests, including molecular genetics, is based on every molecular defect or gene tested i.e. each molecular defect or gene tested is countered as one test.
- 7.3.15 Preanalytical workload (for 'non-test' technical activities) such as media preparation for transport and cell culture for cytogenetic test is captured mainly for the purpose of manpower requirement. It is however, not counted in the total number of tests performed.
- 7.3.16 Any test offered but is not in the list below, requires the workload submission to be temporarily included under 'others' and according to designated groups. Please inform National Pathology Workload Committee via Head of Discipline, of the test names that are to be added into the test list in *Borang Beban Kerja Patologi Bulanan* 1/2016 Hematologi and Borang Beban Kerja Patologi Negeri 1/2016 Hematologi.
- 7.3.17 Workload calculation and recording:

A. GENERAL HAEMATOLOGY

No.	Type of Tests	No. Specimens	Tests Performed	No. Test performed
		No. of	Haemoglobin (automated)	x 1
1	Blood Count	specimens	3 parts Automated Full Blood	x 1
'	Blood Godin	received	Count (FBC) 5 parts Automated Full Blood Count (FBC)	x 1
2	Slide review of abnormal FBC	Not	Slide review of abnormal FBC	x 1
		applicable	Reticulocyte count (Automated)	
3	Reticulocyte count (RC)	specimens	Reticulocyte (Manual)	x 1
4	Erythrocyte Sedimentation Rate		Erythrocyte Sedimentation Rate	x 1
5	G6PD Screening	each test	G6PD Screening	x 1
			Automated Full Blood Count (FBC)	Capture under FBC
_		No. of	Reticulocyte Count	Capture under
6	Full Blood Picture (FBP)	specimens received	Peripheral Blood Film (unstained slide)	RC x 1
			Peripheral Blood Film Morphology	x 1
			Smear /Trephine roll on routine	
7	Bone Marrow Aspiration	No. of specimens received	stains Romanowsky stain : • MGG stains	x 1
			Perls' stains	x 1
	Cytochemical stains :	No. of	Acid Phosphatase	x 1
			α -Naphthol Acetate Esterase	x 1
			Chloracetate Esterase	x 1
			Leucocyte/Neutrophil Alkaline phosphatase (LAP/NAP)	x 1
8		specimens	Myeloperoxidase	x 1
		received	Naphthol AS Acetate Esterase	x 1
			Periodic Acid-Schiff	x 1
			Sudan Black B Stain	x 1
			Tartrate Resistant Acid Phosphatase (TRAP)	x 1
9	Immunohaematology for Klinik Ke	sihatan ONLY		
			Anti-A	x 1
			Anti-B	x 1
			Anti-AB	x 1
	ABO & Rhesus Blood Grouping	No. of specimens	Anti-D	x 1
		received	A Cell	x 1
		received	B Cell	x 1
			O Cell	x 1
10	Others (if test not listed)		Each indivdual test	x 1
	TOTAL		TOTAL	

B. BASIC HEMOSTASIS AND THROMBOSIS

No.	Type of Tests	No. Specimens	Tests Performed	No. Test performed	
1	Activated Partial Thromboplastin Time (APTT)	No. of specimens received	Activated Partial Thromboplastin Time (APTT)	x 1	
	APTT mixing toot (authoritytion	No. of	APTT	Capture under APTT	
2	APTT mixing test (substitution test)	specimens	Initial mixing	x 1	
	test)	received	Additional test : 2 hours incubation mixing	x 1	
			FBC		
	DIVC screening	No. of specimens	Prothrombin Time (PT)	x 1	
3			APTT		
		received	Fibrinogen (activity)		
			FDP/D-Dimer		
4	D-Dimer/FDP		D-Dimer/FDP	x 1	
5	Fibrinogen (activity)		Fibrinogen (activity)	x 1	
6	Prothrombin time (PT/INR)	No. of	Prothrombin time (PT/INR)	x 1	
		specimens	PT	Capture under	
7	PT mixing test (substitution test)	received		PT	
		for each test	Mixing	x 1	
8	Thrombin time (TT)		Thrombin time (TT)	x 1	
9	Others (if test not listed)		Each indivdual test	x 1	
	TOTAL		TOTAL		

C. SPECIALISED HEMOSTASIS AND THROMBOSIS

No.	Type of Tests	No. Specimens	Tests Performed	No. Test performed
1	Activated Protein C Resistance		Activated Protein C Resistance	x 1
2	ADAMTS 13 Activity		ADAMTS 13 Activity	x 1
3	ADAMTS 13 Antigen]	ADAMTS 13 Antigen	x 1
4	Antithrombin	No. of specimens	Antithrombin	x 1
5	Anti-Xa	received for	Anti-Xa	x 1
6	Bleeding Time (BT)	each test	Bleeding Time (BT)	Capture under BT
7	Coagulation Factor V Activity		Coagulation Factor V Activity	x 1
8	Coagulation Factor V Antigen		Coagulation Factor V Antigen	x 1
			PT	Capture under PT
	Coagulation Factor V Assay	No. of specimens received for each test	APTT	Capture under APTT
9			Mixing tests	Capture under mixing test (APTT)
			Factor V Activity	Capture under Factor V Activity
10	Congulation Factor V Inhibitors		PT	Capture under PT
10	Coagulation Factor V Inhibitors		APTT	Capture under APTT

				Capture under
		No. of	Mixing tests	mixing test (APTT)
		specimens received	Dilution in Parallelism study	x 1 for each dilution
		for each test	Dilution in Inhibitor-Bethesda assay	x 1 for each dilution
11	Coagulation Factor VII Activity		Coagulation Factor VII Activity	x 1
12	Coagulation Factor VII Antigen		Coagulation Factor VII Antigen	x 1
			PT	Capture under PT
13	Coagulation Factor VII Assay	No. of specimens	Mixing tests	Capture under mixing test (PT)
		received	Factor VII Activity	Capture under Factor VII Activity
			PT	Capture under PT
14	Coagulation Factor VII Inhibitors	No. of	Mixing tests	Captured under mixing test (PT)
14	Coagulation Factor VII Infilbitors	specimens received	Dilution in Parallelism study	x 1 for each dilution
			Dilution in Inhibitor-Bethesda assay	x 1 for each dilution
15	Coagulation Factor VIII Activity		Coagulation Factor VIII Activity	x 1
16	Coagulation Factor VIII Antigen		Coagulation Factor VIII Antigen	x 1
		No. of specimens received each test	APTT	Capture under APTT
17	Coagulation Factor VIII Assay		Mixing tests	Capture under mixing test (APTT)
			Factor VIII Activity	Capture under Factor VIII Activity
			PT	Capture under PT
18	Coagulation Factor VIII Inhibitors	No. of specimens	Mixing tests	Capture under mixing test (PT)
10	Coagulation Factor vin minibitors	received	Dilution in Parallelism study	x 1 for each dilution
			Dilution in Inhibitor-Bethesda assay	x 1 for each dilution
19	Coagulation Factor IX Activity		Coagulation Factor IX Activity	x 1
20	Coagulation Factor IX Antigen		Coagulation Factor IX Antigen	x 1
		No. of specimens	APTT	Capture under APTT
21	Coagulation Factor IX Assay	received each test	Mixing tests	Capture under mixing test (APTT)
			Factor IX Activity	Capture under Factor IX Activity
		No. of	APTT	Capture under APTT
22	Coagulation Factor IX Inhibitors	specimens received	Mixing tests	Capture under mixing test (APTT)

				x 1 for each
		No. of	Dilution in Parallelism study	dilution
		specimens received	Dilution in Inhibitor-Bethesda assay	x 1 for each dilution
23	Coagulation Factor X Activity	for each test	Coagulation Factor X Activity	x 1
24	Coagulation Factor X Antigen	1	Coagulation Factor X Antigen	x 1
	-		PT	Capture under PT
		No. of	APTT	Capture under APTT
25	Coagulation Factor X Assay	specimens received	Mixing tests	Capture under mixing test (APTT)
			Factor X Activity	Capture under Factor X Activity
			PT	Capture under PT
			APTT	Capture under APTT
26	Coagulation Factor X Inhibitors	No. of specimens received	Mixing tests	Capture under mixing test (APTT)
			Dilution in Parallelism study	x 1 for each dilution
			Dilution in Inhibitor-Bethesda assay	x 1 for each dilution
27	Coagulation Factor XI Activity]	Coagulation Factor XI Activity	x 1
28	Coagulation Factor XI Antigen]	Coagulation Factor XI Antigen	x 1
		No. of specimens received each test	APTT	Capture under APTT
29	Coagulation Factor XI Assay		Mixing tests	Capture under mixing test (APTT)
			Factor XI Activity	Capture under Factor XI Activity
			APTT	Capture under APTT
30	Coagulation Factor XI Inhibitors	No. of specimens	Mixing tests	Capture under mixing test (APTT)
		received	Dilution in Parallelism study	x 1 for each dilution
			Dilution in Inhibitor-Bethesda assay	x 1 for each dilution
31	Coagulation Factor XII Activity		Coagulation Factor XII Activity	x 1
32	Coagulation Factor XII Antigen		Coagulation Factor XII Antigen	x 1
		No. of	APTT	Capture under APTT
33	Coagulation Factor XII Assay	specimens received each test	Mixing tests	Capture under mixing test (APTT)
			Factor XII Activity	Capture under Factor XII Activity
34	Coagulation Factor XII Inhibitors		APTT	Capture under APTT

				Capture under
		No. of	Mixing tests	mixing test (APTT)
		specimens received	Dilution in Parallelism study	x 1 for each dilution
		each test	Dilution in Inhibitor-Bethesda assay	x 1 for each dilution
35	Coagulation Factor XIII Activity	-	Coagulation Factor XIII Activity	x 1
36	Coagulation Factor XIII Antigen		Coagulation Factor XIII Antigen	x 1
			PT	Capture under PT
37	Coagulation Factor XIII Inhibitors	No. of specimens	APTT	Capture under APTT
37	Coagulation Factor All Illinoitors	received	Dilution in Parallelism study	x 1 for each dilution
			Dilution in Inhibitor-Bethesda assay	x 1 for each dilution
38	Coagulation Factor XIII Screening	No. of specimens received	Coagulation Factor XIII Clot Lysis Test/Screening	x 1
			Bleeding Time	Capture under BT
		No. of specimens received	FBC (Hb & Platelet count)	Capture under FBC
	Coagulation profile		Fibrinogen	Capture under Fibrinogen
39			PT	Capture under PT
			APTT	Capture under APTT
			Thrombin Time	Capture under TT
			D-Dimer	Capture under D-Dimer
40	Coagulation Tissue factor		Coagulation Tissue Factor	x 1
41	Euglobulin Clot Lysis		Euglobulin Clot Lysis	x 1
42	Fibrin and Fibrinogen degradation product (FDP)		Fibrin and Fibrinogen degradation product (FDP)	x 1
43	Fibrin Monomer	No. of	Fibrin Monomer	x 1
44	Fibrinogen Antigen	specimens	Fibrinogen Antigen	x 1
45	Fibrinogen Activity	received	Fibrinogen Activity	x 1
		for each test	Platelet Factor 4	x 1
46	Heparin induced thrombocytopaenia test		Serotonin Assay	x 1
	un em Beeytepaerna teet		Platelet Aggregation for HIT	x 1 per agonist
47	Kininogen High Molecular Weight		Kininogen High Molecular Weight	x 1
			Prothrombin Time	Capture under PT
40	L	No. of	APTT (LA Sensitive)	x 1
48	Lupus anticoagulant screen	specimens received	DRVVT Screening	x 1
		10001100	Silica Clotting Time screen	x 1
			Kaolin Clotting Time	x 1

			DRVVT Confirm	v. 1
		N/ 5		x 1
49	Lupus anticoagulant confirm	No. of specimens	Platelet Neutralization Test (PNT)	x 1
49	Lupus anticoagulant commi	received	Silica Clotting Time confirm	x 1
		70007704	Hexagonal Phosphatidyl etanolamin (HPE)	x 1
50	Plasmin Inhibitor		Plasmin Inhibitor	x 1
51	Plasminogen	No. of	Plasminogen	x 1
52	Plasminogen Activator Inhibitor 1 (PAI 1)	specimens received	Plasminogen Activator Inhibitor 1 (PAI 1)	x 1
53	Platelet aggregation test	for each test	Platelet aggregation test (Agonists : ADP/Collagen etc)	x 1 per agonist
			Bleeding Time	Capture under BT
			Clot Retraction	x 1
			Platelet Marker - 1 test per MoAb used :	
			• CD 41a	x 1
		No. of	• CD 42b	x 1
54	Platelet function tests	specimens received	Platelet aggregation test: 1 test per agonist used	
			• ADP	x 1
			Collagen	x 1
			Ristocetin	x 1
			Arachidonic acid	x 1
			Epinephrine	x 1
55	Platelet Factor 3		Platelet Factor 3	x 1
56	Platelet Factor 4		Platelet Factor 4	x 1
57	Platelet Function Analysis (PFA 100)		Platelet Function Analysis (PFA 100)	x 1 per marker
58	Platelet Antibody		Platelet Antibody	x 1
59	Protein C Activity		Protein C Activity	x 1
60	Protein C Antigen	No. of	Protein C Antigen	x 1
61	Protein S Antigen	specimens	Protein S Antigen	x 1
62	Protein S, Free	received for each test	Protein S, Free	x 1
63	Protein S, Total	101 00011 1001	Protein S, Total	x 1
64	Prothrombin Antigen		Prothrombin Antigen	x 1
65	Prothrombin Fragment 1& 2		Prothrombin Fragment 1& 2	x 1
66	Reptilase Test		Reptilase Test	x 1
67	Thromboglobulin, Beta		Thromboglobulin, Beta	x 1
68	Thrombomodulin		Thrombomodulin	x 1
			Lupus Anticoagulant	Capture under Lupus Anticoagulant
			Antithrombin Antigen	x 1
			Antithrombin Activity	x 1
69	Thrombophilia screening	No. of specimens	Protein C Activity	Capture under Protein C Activity
		received	Protein S, Total	Capture under Protein S, Total
			Protein S, Free	Capture under Protein S, Free
			Activated Protein C Resistance with Factor V (Normal Plasma)	x 1

				Capture under
			Factor VIII : C Activity	Factor VIII Activity
			Plasma Homocysteine Level	x 1
			Heparin Cofactor II	x 1
			Plasminogen Level	x 1
			Fibrinogen	Capture under Fibrinogen
			D-Dimer	Capture under D-Dimer
70	von Willebrand Ristocetin Cofactor assay	No. of specimens	von Willebrand Ristocetin Cofactor assay	x 1
71	von Willebrand Factor (vWF) Activity	received for each test	von Willebrand Factor (vWF) Activity	x 1
72	von Willebrand Factor (vWF) Antigen	No. of	von Willebrand Factor (vWF) Antigen	x 1
73	vWF collagen binding assay	specimens received for each test	vWF collagen binding assay	x 1
74	von Willebrand Multimers Analysis		von Willebrand Multimers Analysis	x 1
			APTT	Capture under APTT
			vWF: Ag	Capture under vWF: Ag
			vWF: Activity	Capture under vWF: Activity
			Coagulation Factor VIII Activity	x 1
75	von Willebrand disease profile	No. of specimens	vWF:CB	x 1
	Ten Trinestana aleeade preme	received	vWF:Rco	x 1
			Additional tests (Subtyping):	
			vWF Factor VIII Binding assay	x 1
			Ristocetin Induced platelet agglutination (RIPA)	x 1
			Multimeric analysis	x 1
76	Warfarin Level	No. of specimens	Warfarin Level	x 1
77	Others (if test not listed)	received for each test	Each indivdual test	x 1
	TOTAL		TOTAL	

D. RED CELL DISORDERS

No.	Type of Tests	No. Specimens	Tests Performed	No. Test performed
1	Autohaemolysis test		Autohaemolysis test	x 1
2	G6PD Activity		G6PD Activity	x 1
3	Heinz Bodies	No. of	Heinz Bodies	x 1
4	Ham test	specimens	Ham test	x 1
5	Methaemoglobin reduction test	received for	Methaemoglobin reduction test	x 1
6	Osmotic Fragility Test		Osmotic Fragility Test	x 1
7	Pyruvate Kinase Activity	each test	Pyruvate Kinase Activity	x 1
8	Schumm test		Schumm test	x 1
9	Sucrose Lysis Test		Sucrose Lysis Test	x 1
10	Others (if test not listed)	No. of specimens received	Each indivdual test	x 1
	TOTAL		TOTAL	

E. HAEMOGLOBIN DISORDERS

No.	Type of Tests	No. Specimens	Tests Performed	No. Test performed
			Automated Blood Count	Capture under FBC
			Reticulocyte count	Capture under reticulocyte count
	Hb Analysis (Basic tests)	No. of specimens received	Peripheral Blood Film (unstained slide)	Capture under peripheral blood film (unstained slide)
1			Peripheral Blood Film Morphology	x 1
'			Hb quantification (HPLC or Hb Capillary electrophoresis)	x 1
			H inclusion	x 1
			HPLC	x 1
		No. of specimens	Hb Capillary Electrophoresis	x 1
	Hb Analysis (Additional tests)		Hb Electrophoresis (alkaline)	x 1
		received	Hb Electrophoresis (acid)	x 1
2	Haemogloblin, Urine		Haemogloblin, Urine	x 1
3	Haemogloblin Fetal (Kleihauer test)	No. of	Haemogloblin Fetal (Kleihauer test)	x 1
4	Haemoglobin, Thermolabile (Unstable Hb)	specimens received for each test	Haemoglobin, Thermolabile (Unstable Hb)	x 1
5	Sickling Test		Sickling Test	x 1
6	Others (if test not listed)		Each indivdual test	x 1
	TOTAL		TOTAL	

F. IMMUNOPHENOTYPING

1 Fetal red cells (HbF) quantitation 2 Cerebral spinal fluid (CSF) for blasts No. of specimens received No. of specimens received No. of specimens received Fetal red cells (HbF) quantitation CSF cytospin Immunophenotyping FBC Blood film Bone Marrow Film	x 1 X MoAb used Capture under FBC x 1
2 Cerebral spinal fluid (CSF) for blasts specimens received Immunophenotyping FBC Blood film	X MoAb used Capture under FBC x 1
blasts specimens received Immunophenotyping FBC Blood film	Capture under FBC x 1
FBC Blood film	FBC x 1
Acute Leukaemia Rone Marrow Film	
	x 1
Immunophenotyping : Basic CSF cytospin	x 1
screening Body Fluid cytospin	x 1
No. of Acute Leukemia screening	g (SA) x MoAb used
3 specimens received Cell Viability	x 1
Myeloperoxidase (MPO)	Capture under MPO
Acute leukaemia B Cell - Acute Lymphobla immunophenotyping : Leukemia (B-ALL)	stic x MoAb used
Diagnostic panel T Cell - Acute Lymphobla: Leukemia (T-ALL)	stic x MoAb used
Acute Myeloblastic Leuke	emia (AML) x MoAb used
FBC	Capture under FBC
Lymphoproliferative disorder/	x 1
Lymphoma Bone Marrow Film	x 1
Immunophenotyping: Basic	x 1
screening No. of Body Fluid cytospin	x 1
4 specimens LPD/ Lymphoma screening received	ng (SL) x MoAb used
Cell Viability	x 1
Lymphoproliferative disorder/ B - Cell Lymphoproliferative (B-LPD)	ve Disorder x MoAb used
Lymphoma Immunophenotyping: Diagnostic panel T - Cell Lymphoproliferativ (T-LPD)	ve Disorder x MoAb used
Natural Killer Cell (NK)	x MoAb used
FBC	Capture under FBC
Multiple Myeloma Blood film	x 1
Immunophenotyping :Basic Screening No. of Screening No. of Screening	x 1
5 Specimens Multiple Myeloma Screenii	ng (SM) x MoAb used
received Cell Viability	x 1
Multiple Myeloma Immunophenotyping: Additional Multiple Myeloma (MM) paragraphical	anel x MoAb used
FBC	Capture under FBC
Residual disease of acute No. of Ieukaemia Blood film	x 1
b Immunophonotyping Specimens Bone Marrow Film	x 1
(for B-ALL) received B-ALL MRD panel	x MoAb used
Cell Viability	x 1

	Decided discours of costs	No. of specimens received	FBC	Capture under FBC
	Residual disease of acute leukaemia		Blood film	x 1
7	Immunophenotyping		Bone Marrow Film	x 1
	(for T-ALL)	received	T-ALL MRD panel	x MoAb used
			Cell Viability	x 1
	Residual disease of acute		FBC	Capture under FBC
	leukaemia	No. of	Blood film	x 1
8	Immunophenotyping	specimens received	Bone Marrow Film	x 1
	(for AML)	received	AML MRD panel	x MoAb used
			Cell Viability	x 1
	CD4/CD8 Count (Single platform)	No. of specimens received	CD3/4/8/45	x MoAb used
9	CD4/CD8 Count (Dual	No. of specimens	FBC	Capture under FBC
	platform)	received	CD3/4/8/45	x MoAb used
10	Neutrophil activation	No. of specimens received	e.g.CD64	x MoAb used
11	Platelets Glycoproteins Immunophenotyping	No. of specimens received	As per number of monoclonal antibodies (MoAb) used	x MoAb used
	Paroxysmal nocturnal	No. of	FBC	Capture under FBC
12	haemoglobinuria (PNH) Immunophenotyping	specimens received	PNH Clone in White cells	x MoAb used
	minunoprieriotyping		PNH Clone in red cells	x MoAb used
13	Others (if test not listed)	No. of specimens received	Each indivdual test	x 1
	TOTAL		TOTAL	

G. MOLECULAR DIAGNOSIS FOR NON MALIGNANT HAEMATOLOGY

Method Classification*:

M: Manual

S: Semi automation F: Full automation

No.	Type of Tests	No. Spe	cimens	Tests Pe	rformed (M / S / F)*	No. Test performed	
1	Molecular tests for G6PD genotyping	No. of specimens received		As per type of molecular defect tested e.g. Viangchan variant Canton variant Mahidol variant Kaiping variant		x Molecular defect tested	
2	Molecular tests for Haemophilia		No. of specimens received		molecular defect tested ersion ersion	x Molecular defect tested	
3	Molecular tests for	Thalassaemia	/Haemoglobi	inopathy			
3.1	Molecular tests for Beta-globin	ular tests No. of		mutation)	p-PCR (deletional FH-3, Chinese, Asian)	x Molecular defect tested	
	-	gene - 11 parts	9 - 11	-88 [C>T] (β ⁺)			
					-86 [C>G] (β ⁺)		
					-29 [A>G] (β ⁺)		
					-28 [A>G] (β ⁺)		
					Cap+1 [A>C] (β ⁺)		
					Initiation codon [A <i>T</i> G>A <i>G</i> G] (β°)		
						Codon 8/9 [+G] (β°)	
				β Multiplex	Codon 15 [TGG>TAG] (β°)		
				ARMS-PCR e.g.	Codon 16 [GGC>GG-] (β°)	x Molecular defect tested	
					Codon 17 [AAG>TAG]		
					(β°)		
					Codon 19 [AAC>AGC] Malay (β^{+})		
					Codon 26		
					[$GAG>AAG$] Hb E (β ^E)		
					IVS 1-1 [G>T] (β°)		
					IVS 1-1 [G>A] (β°)		
					IVS 1-5 [G>C] (β ⁺)		

					Codon 41/42 [-TTCT] (β °) Codon 43[GAG>TAG](β °) Codon 71/72 [+A] (β °) IVS 2-654 [C>T] (β [†]) Poly A [AATAAA >AATAGA] (β [†]) 619bp deletion	
				β FIL deletion		x Molecular
				Singleplex β se	equencing PCR	defect tested
					PCO7_F	
					PCO_R	
				β Sequencing	833_1338	x Molecular
				(4 reactions /sample)	PAA_R	defect tested
				, campio,	CA	
					Common F	
				O MI DA	B minus 296_Fw	
				β MLPA		
					B) Multiplex Gap-PCR	
				Asian-Indian De Gap PCR	el/Inv $^{\rm G}$ γ($^{\rm A}$ γδβ) Multiplex-	
				Hb Lepore Mult	iplex Gap-PCR	
				β Multiplex ARI	MS-PCR for HbS	x Molecular
				β Multiplex ARN Punjab	MS-PCR for HbD	defect tested
				β Multiplex ARI	MS-PCR for HbC	
					re Variant (Hb Lepore epore Baltimore, Hb gton-Boston)	
				<u> </u>	hism (RFLP-PCR)	
3.2	Molecular tests for Alpha globin gene defects	No. of specimens received	Alpha globin gene – 6 parts	α Multiplex Gap-PCR	Single gene deletion $-\alpha^{3.7}$ Single gene deletion $-\alpha^{4.2}$ Double genes	x Molecular
			(deletional mutation) e.g	deletion ^{SEA} Double genes deletion ^{FIL} Double genes deletion ^{THAI}	defect tested	

				α Multiplex ARMS-PCR e.g.	Double genes deletion ^{MED} Double genes deletion(□ ^{20.5} Initiation codon (ATG>A_G) Codon 30 (ΔGAG) Codon 35 (TCC→CCC) Codon 59 (GGC→GAC) Codon 125 (CTG→CCG) or Hb Quong Sze Termination codon	x Molecular defect tested
				Singleplex α seq and HBA2 gene)	(TAA→CAA) or Hb constant spring uencing PCR (HBA1	x Molecular defect tested
				and the gener	AC40_R	
					BE17_R	
				α sequencing	BE10_F	
				(covers α1 and	AD F	x Molecular
				α2 gene; 8 reactions/	AC40_R	defect tested
				sample)	BE12_R	
					BE10_R	
					AD_F	
				α MLPA		
				α triplication PCF	?	x Molecular
						defect tested
		A/- C	A	·	S-PCR for Hb Pakse	
4	Molecular tests for Thrombophilia	No. of specimens received	FV Leide	of molecular defe en mutation 0A mutation	ct tested e.g.	x Molecular defect tested
5	Molecular tests for von Willebrand disease	No. of specimens received	Single cNonsens	of molecular defe ytosine deletion in se mutation in exo se mutation in exo	exon 18 ns 28	x Molecular defect tested
6	Others (if test not listed)	No. of specimens received	Each indivdual test			x Molecular defect tested
	TOTAL				TOTAL	
<u> </u>	l		i			

H. MOLECULAR DIAGNOSIS FOR MALIGNANT HAEMATOLOGY

Method Classification*:

M: Manual

S: Semi automation F: Full automation

No.	Type of Tests	No. Specimens	Tests	Performed (M / S / F)*	No. Test performed	
	Molecular tests for		RNA quantificati	RNA quantification		
1	leukaemia : Screening	No. of specimens	Master Screenir	ng	x Molecular defect tested	
	Molecular tests for leukaemia : Confimation	received	Split Out Analys	is	x Molecular defect tested	
2	Acute myeloid leukaemia mutation studies	No. of specimens received	AML mutation studies (PCR & Sequencing) e.g.	FLT-3 NPM1 CEBPA c-KIT CBF /MYH11 RUNX1/RUNX1T1	x Molecular defect tested	
3	Acute lymphoid leukaemia mutation studies	No. of specimens received	ALL mutation studies (PCR & Sequencing) e.g.		x Molecular defect tested	
4	Molecular tests for Lymphoma	No. of specimens received	As per type of molecular defect to be detected e.g. BCL2 rearrangement MYC translocation		x Molecular defect tested	
	Myeloproliferative Neoplasm Mutation	No. of	DNA quantificati	ion	x 1	
	Sudy: Basic screening	specimens received	JAK2V617F (JA	K2 Mutation)	x Molecular defect tested	
5	Myeloproliferative	., .	JAK 2 exon 12			
	Neoplasm Mutation Study: Additional	No. of specimens	MPL W515L/K		x Molecular defect tested	
	tests	received	CALR			
		No. of	FBC		Capture under FBC	
6	PML RARA qualitative	specimens received	RNA quantificati	ion	x 1	
	•	received	PML-RARA qu	alitative	x Molecular defect tested	
		No. of	FBC		Captured under FBC	
7	PML RARA quantitative	specimens received	RNA quantificati	ion	1 test	
		received	PML-RARA qu	x Molecular defect tested		

		No. of	FBC	Capture under FBC
8	BCR-ABL qualitative	specimens	RNA quantification	1 test
		received	BCR-ABL qualitative	x molecular defect tested
		No. of	FBC	Captured under FBC
9	BCR-ABL quantitative	specimens received	RNA quantification	x 1
		received	BCR-ABL quantification	x Molecular defect tested
40	Chronic myeloid	No. of	DNA quantification	x 1
10	leukaemia mutation studies	specimens received	CML mutation studies e.g T315I mutation detection	x Molecular defect tested
		No. of specimens received	FBC	Captured under FBC
11	MRD for B-ALLL		RNA quantification	x 1
			Specific marker quantification	x Molecular defect tested
		No. of	FBC	Captured under FBC
12	MRD for T-ALLL	specimens	RNA quantification	x 1
		received	Specific marker quantification	x Molecular defect tested
		No. of	FBC	Captured under FBC
13	MRD for AML	specimens	RNA quantification	x 1
		received	Specific marker quantification	x Molecular defect tested
14	Others (if test not listed)	No. of specimens received	Each indivdual test	x Molecular defect tested
	TOTAL		TOTAL	

I. GENETICS FOR HAEMATOLOGICAL DISORDER

Method Classification*:

M: Manual

S: Semi automation F: Full automation

No.	Type of Tests	No. Specimens	Tests Performed (M / S / F)*	No. Test performed
1	Chromosomal microarray	No .of	Array CGH analysis	x 1
2	Chromosome Analysis	specimens received	Chromosome Analysis	x 1
3	Chromosome Breakages, Fanconi's Anaemia	for each tests	Chromosome Breakages, Fanconi's Anaemia	x 1

4	Genetic linkage analysis for Haemophilia A	No .of specimens received	As per marker used e.g Hind III ST 14 CA13 VNTR CA22 VNTR	x Molecular defect tested
5	Molecular genetic BCR-ABL	No .of specimens	Fluorescence-in-situ hybridization (FISH), BCR-ABL	x 1
6	Molecular genetic RARA	received for each	Fluorescence-in-situ hybridization (FISH), RARA	x 1
7	Molecular genetic PML-RARA	tests	Fluorescence-in-situ hybridization (FISH), PML/RARA	x 1
8	Molecular genetic ETV6-RUNX1	No. of specimens received	Fluorescence-in-situ hybridization (FISH), ETV6-RUNX1	x 1
9	Others (if test not listed)	No. of specimens received	Each indivdual test	x 1
	TOTAL		TOTAL	

J. STEM CELL TRANSPLANTATION

No.	Type of Tests	No. Specimens	Tests Performed	No. Test performed
1	Cell Viability	No. of specimens received	Cell Viability	x 1
2	CD 34 count of bone	No. of	FBC	Capture under FBC
2	marrow, PBSC, cord blood		CD34	x 1
		received	Cell Viability	x 1
	CD 34 count for tranplant		FBC	Capture under FBC
	without processing (bone marrow /PBSC/Donor lymphocyte)	No. of specimens received	Blood film morphology	x 1
3			CD34	x 1
			CD3 Cells	x 1
			Cell viability	x 1
			FBC pre processing	Capture under FBC
			No. of specimens received CD34 Cell Viability FBC Blood film morphology CD34 CD3 Cells Cell Viability	Capture under FBC
	CD 34 count for stem cell	No. of	Blood film morphology	x 1
4	processing (bone marrow/ PBSC/ cord blood/Donor	specimens	CD34 pre processing	x 1
	lymphocyte)	received	CD34 post processing	x 1
	lymphocyte)		CD3 Cells pre processing	x 1
			CD3 Cells post processing	x 1
			Cell viability pre processing	x 1
			Cell viability post processing	x 1

			Culture & sensitivity test pre & post processing	Capture under Microbiology
			Stem cell Cryopreservation	x 1
			Donor Lymphocyte Cryopreservation	x 1
	Additional Stem cell	No. of	Red Cell Depletion Procedure	x 1
	processing or test	specimens received	Stem Cell (CD34) Selection Procedure	x 1
			T Cell Depletion Procedure	x 1
			HLA Typing of cord blood	Workload capture by referral labs (IMR/PDN)
	Infusion of cryopreserved		Infusion of cryopreserved stem cell	x 1
5	stem cell/ donor lymphocytes	No. of specimens received for each test	Infusion of cryopreserved donor lymphocytes	x 1
6	CFU for stem cell		CFU for stem cell	x 1
	Chimerism studies (STR) : Diagnostic panel		Donor allele identification	x Molecular markers tested
7			Pre-Recipient alelle identification	x Molecular markers tested
			1st Post-Recipient STR asessment	x Molecular markers tested
8	Chimerism studies (STR):	, , , , , , , , , , , , , , , , , , , ,	1st Post-Recipient STR asessment	x Molecular markers tested
	Follow up panel		FBC	Captured under FBC
9	Chimerism studies		Real-time PCR – Genotyping	x Molecular markers tested
ש	(Real-time PCR)		Real-time PCR - Quantitation of chimerism	x Molecular markers tested
10	Others (if test not listed)	No. of specimens received	Each indivdual test	x Molecular markers tested
	TOTAL		TOTAL	

PREANALYTICAL WORKLOAD (FOR 'NON-TEST' TECHNICAL ACTIVITIES)

Activity	Workload
Cytogenetic Cell culture	Number of test tubes
Cytogenetic transport media	Number of test tubes
Cytogenetic slide preparation	Number of slides

PATHOLOGY SERVICES MINISTRY OF HEALTH, MALAYSIA

REPORT ON LABORATORY WORKLOAD: HAEMATOLOGY

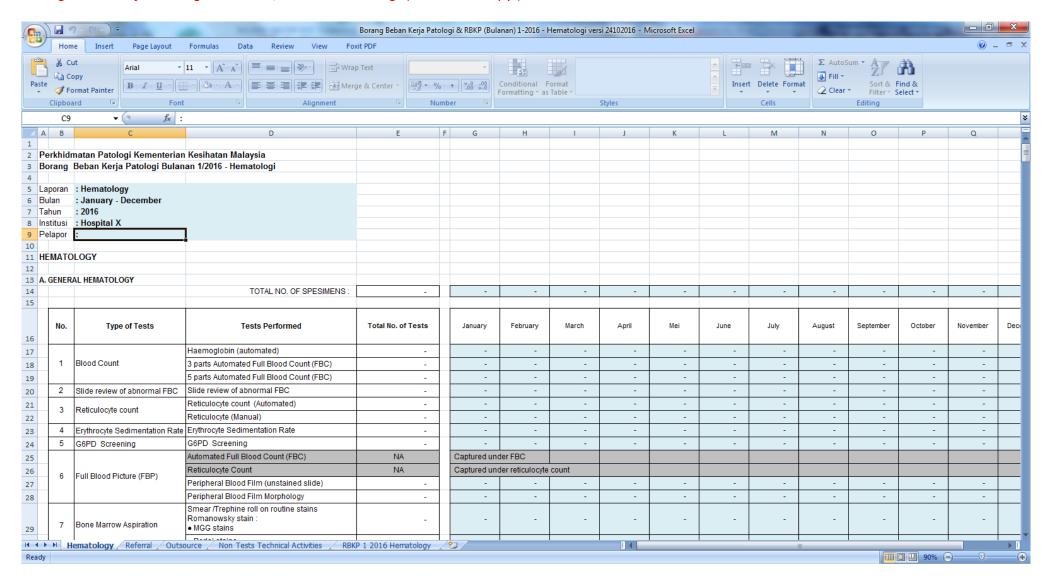
FOR THE OF MONTH:	YEAR :

Month or Location of Hospitals, Institutions and Public Health Facilities	General Haematology		Haemostasis & Thrombosis				Red Cell		Haemoglobin		Immuno		Molecular Diagnosis				Genetics for		Stem Cell			
			Basic		Specialised		Disorder		Disorder		phenotyping		Non Malignant Haematology		Malignant Haematology		Heamatological disorder		Transplantation		Total No. of Specimens	Total No. of Tests
	S	Т	S	Т	S	Т	S	Т	S	Т	S	Т	S	Т	S	Т	S	Т	S	Т	Tota Spe	Spe Tota Test

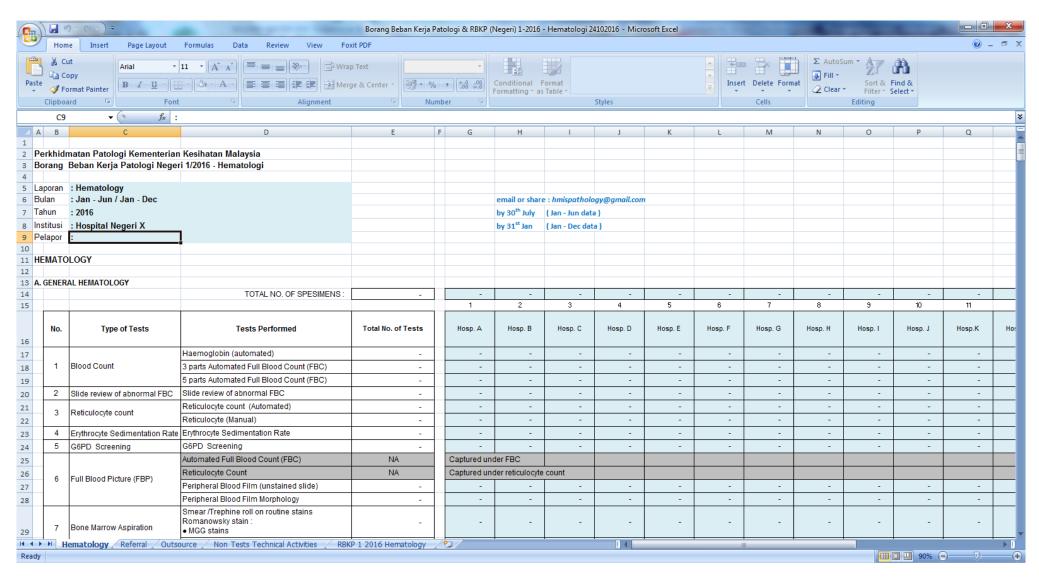
Page 1 of 1

S=Number of Spesimen T=Number of Tests

Borang Beban Kerja Patologi Bulanan 1/2016 - Hematologi ('Excel Soft Copy')



Borang Beban Kerja Patologi Negeri 1/2016 - Hematologi ('Excel Soft Copy')



7.4 MEDICAL MICROBIOLOGY

- 7.4.1 In addition to workload reporting to MOH using PER-SS 206 (Pin. 1/2000) form, hospital laboratories are also to submit to their respective State Pathologists, in soft copy, detailed (granular), as well as, summarised workload data on Medical Microbiology, using Borang Beban Kerja Patologi Bulanan 1/2016 Mikrobiologi Perubatan and Borang Ringkasan Beban Kerja Patologi 1/2016 Mikrobiologi Perubatan, respectively.
- 7.4.2 The total number of specimens received by a laboratory is calculated from total number of specimens received across the designated groups (bacteriology, mycology, parasitology, immunology and virology).
- 7.4.3 Culture is divided into primary culture (culture from original sample including blood culture bottle) and secondary culture (second culture done from primary culture/broth e.g. culture from positive bottle; culture done from Selenite F). Myco/F lytic bottle is included in the blood culture in bacteriology section. The number of culture is counted as one regardless of the number of media plates used.
- 7.4.4 Biochemical identification is divided into preliminary and definitive identification and also according to test methods e.g manual (M), commercial kit (C) or automation (A). Example of preliminary biochemical testing is screening of stool pathogens e.g Enteropathogenic E coli (EPEC), Salmonella and Shigella. The number of test for biochemical identification is counted as one test for one set of biochemical identification done per organism, regardless of the number of biochemicals and methodology used.
- 7.4.5 Serotyping includes Lancefield grouping and serotyping for organisms such as *Salmonella, Shigella, Vibrio*, EPEC, *Haemophilus* etc. For hospitals, the number of test will be counted as one test for each organism and will be included in the culture workload. For referral laboratories e.g IMR and MKAK which performed surveillance and outbreak investigations, the number of test will be the number of actual serotyping done.
- 7.4.6 Antimicrobial sensitivity testing are divided into manual (disk diffusion, which may be further divided into preliminary and definitive), automated (e.g Vitek) or on a single plate e.g yeast broth sensititer. Each test is counted as one regardless of the number of plates or strips used. For institution which practices preliminary antimicrobial testing from blood culture bottle, the procedure is counted as one test.
- 7.4.7 Detection of multidrug resistance e.g ESBL confirmation test, modified Hodge test is counted under culture and sensitivity for hospitals as the test is done on the same specimen for culture.
- 7.4.8 Each profile testing in serology/immunology using a multitest kit is counted as one test e.g. several antibodies/antigens are detected on a single slide, cartridge or plate. If an individual test kit is used for that profile, the number of test is counted as one for each test. For antibody testing, the number of tests for different class of antibody are counted separately.

- 7.4.9 Any subsequent dilution performed following a screening test e.g. ANA and RPR, is counted as one additional test, regardless of the number of serial dilutions performed.
- 7.4.10 For molecular testing, the test is classified according to method of testing i.e. manual, semi automation or full automation. Regardless of the method of testing, the workload calculation is based on every gene tested i.e. each gene tested is considered as one test.
- 7.4.11 There are several tests generally performed under the umbrella of Chemical Pathology that are also run in Medical Microbiology / Serology / Immunology laboratories. These tests include urine biochemistry (striptest/dipstick-Qualitative), urine pregnancy test, urine microscopy (manual or automated), CRP, C3, C4, IgG, IgA and IgM and total IgE. The workload for these tests are captured depending on the local practices and the discipline performing the tests. The final workload however, is encouraged to be reported under Chemical Pathology. In T & B Lymphocyte Subset Enumeration (Dual platform method) test, FBC is to be reported under cytology and for seminal fluid analysis, the number of tests will be reported under cytology.
- 7.4.12 Preanalytical workload (for 'non test technical activities') such as media preparation, as well as slide preparation for TB EQA and Indirect Immunoperoxidase (IIP) test is captured mainly for the purpose of manpower requirement. It is however, not counted in the total number of tests performed.
- 7.4.13 Any test offered that is not in the list, requires the workload submission to be temporarily added under 'others' and according to designated groups. Please inform National Pathology Workload Committee via Head of Discipline, of the test names that are to be added into the test list in both *Borang Beban Kerja Patologi Bulanan 1/2016 Mikrobiologi* and *Borang Beban Kerja Patologi Negeri 1/2016 Mikrobiologi*.
- 7.4.14 Workload calculation and recording:

A. BACTERIOLOGY

No.	Type of Tests	No. Specimens	Tests Performed	No. Test performed		
			Wet mount	x 1		
A1	Microscopy	No. of specimens	Gram stain	x 1		
		received for each	Indian ink	x 1		
A2	Cell count	test	Cell count	x 1		
A3	Bacterial antigen detection		Bacterial antigen test	x 1		
	Culture (all types of specimen)		Primary culture	x 1		
	Culture (all types of specimen)		Secondary culture	x 1		
		No. of specimens	Preliminary biochemical identification	x 1		
A4	Biochemical Identification	received	Biochemical identification (Manual)	x 1		
	Biochemical identification	70007704	Biochemical identification	x 1		
			(Commercial e.g API)	^ /		
			Biochemical identification (Automation e.g Vitek)	x 1		

	Serotyping		Serotyping	x 1
			Preliminary Antibiotic Sensitivity	x 1
			testing (Disk diffusion)	ΧΙ
	Antibiotic sensitivity testing		Antibiotic Sensitivity Testing (Disk diffusion)	x 1
	, ,		Antibiotic Sensitivity Testing (E test)	x 1
		No. of specimens received	Antibiotic sensitivity testing (Automation e.g Vitek)	x 1
		, , , , , , , , , , , , , , , , , , , ,	AmpC	x 1
			B- lactamase	x 1
	Detection of resistance		ESBL	x 1
			Modified Hodge test	x 1
			MRSA MecA	x 1
		No. of specimens	Direct smear (Ziehl Neelsen)	x 1
		received	Direct smear (IF)	x 1
		No. of specimens	Conventional culture	x 1
		received	Automated culture	x 1
		No. of specimens	MTB - conventional PCR	x 1
A5	TB (all types of specimens)	received	MTB - fully automated PCR	x 1
		No. of specimens	MDRTB LPA	x 1
		received	MDRTB GeneXpert	x 1
		No. of specimens	Antibiotic sensitiity testing - first line	x 1
		received	Antibiotic sensitiity testing - second line	x 1
			Slit skin smear	x 1
		No. of specimens	Culture	x 1
A6	Mycobacterium leprae	received for each	Conventional PCR	x 1
		tests	Fully automated PCR	x 1
			AST	x 1
			AmpC	x 1
			B lactamase	x 1
			ESBL	x 1
	Detection of multidrug resistance		Modified Hodge test	x 1
A7	organisms/resistance gene	No. of specimens received for each	CaMRSA - MecA gene,	x 1
A	(for referral laboratories)	tests	CaMRSA - PVL gene	x 1
			KPC	x 1
			OXA	x 1
			VIM	x 1
			IMP	x 1
			Haemophilus influenzae serotyping	
A8			E coli serotyping serotyping	x actual
	Sorotyping	No. of specimens	Salmonella serotyping	number of
	Serotyping (for referral laboratories)	received for each	Shigella serotyping	serotyping performed
	(1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	organism	Vibrio cholerae serotyping	for each
			Strep pneumoniae serotyping	organism
			Neisseria meningitidis serotyping	
A9	Serology			
1	Anti- Streptolysin O titre (ASOT)	No. of specimens	Anti- Streptolysin O titre	x 1
2	Bartonella henselae Antibody	received for	Bartonella henselae IgG	x 1
-		each test	Bartonella henselae IgM	x 1

3	Borrelia burgdorferi Antibody		Borrelia burgdorferi IgG	x 1
			Borrelia burgdorferi IgM	x 1
4	Brucella abortus Antibody		Brucella abortus IgG	x 1
7	Bracona abortae runibody		Brucella abortus IgM	x 1
_	Drugalla malitanaia Antibadu		Brucella melitensis IgG	x 1
5	Brucella melitensis Antibody	No of anasimona	Brucella militensis IgM	x 1
_		No. of specimens received for	Chlamydia pneumoniae IgG	x 1
6	Chlamydia pneumoniae Antibody	each test	Chlamydia pneumoniae IgM	x 1
_		Cacri test	Chlamydia psittaci IgG	x 1
7	Chlamydia psittaci Antibody		Chlamydia psittaci IgM	x 1
•			Chlamydia trachomatis IgG	x 1
8	Chlamydia trachomatis Antibody		Chlamydia trachomatis IgM	x 1
9	Clostridium difficile toxin assay		Clostridium difficile toxin assay	x 1
40	•		Coxiella burnetii IgG	x 1
10	Coxiella burnetii Antibody		Coxiella burnetii Ig M	x 1
	005.1/551	No. of specimen	CSF VDRL screening	x 1
11	CSF VDRL	received	CSF VDRL dilution	x 1
		No. of specimens	Indirect Immunoperoxidase (IIP)	x 1
12	Rickettsial Antibody	received for	Rickettsial IgG	
	•	each test	Indirect Immunoperoxidase (IIP) Rickettsial IgM	x 1
		No. of specimens	Legionella pneumophilia Ag	x 1
13	Legionella pneumophilia serology	received for	Legionella pneumophilia IgG	x 1
		each test	Legionella pneumophilia IgM	x 1
	Leptospira Antibody		Leptospira IgM	x 1
14		No. of specimens received for	Leptospira IgG	x 1
17		each test	Lepto spira MAT	x 1
		040711001	Lepto spira MAT dilution	x 1
15	Melioidosis IgM	No. of specimen	Melioidosis IgM	x 1
13	Welloidosis IgiVi	received	Melioidosis IgM dilution	x 1
16	Mycoplasma pneumoniae		Mycoplasma pneumoniae IgG	x 1
	Antibody		Mycoplasma pneumoniae IgM	x 1
17	Atypical pneumonia screening (Pneumobact) IF	No of anasimana	Atypical pneumonia screening (Pneumobact) IF	x 1
18	Strep pneumoniae urinary antigen	No. of specimens received for	Strep pneumoniae urinary antigen	x 1
10	Streptococcus, Group B urinary	each test	Streptococcus, Group B urinary	v 1
19	antigen		antigen	x 1
20	TPHA or TPPA		TPHA or TPPA	x 1
21	Typhoid Antibody		Typhidot IgG	x 1
- '	. , , , , , , , , , , , , , , , , , , ,		Typhidot IgM	x 1
22	RPR/VDRL Serum	No. of specimens	RPR /VDRL screening	x 1
		received	RPR/VDRL dilution	x 1
A10	Molecular			
1	Bordetella pertussis	No. of specimens received for each	Bordetella pertussis	x 1
2	Brucella	tests	Brucella	x 1
	Identification of bacteria by	No. of specimens	16s sequencing Forward sequencing	x 1
3	16sRNA	received	16s sequencing Reverse	x 1
	10011111			
	100111111	No. of specimens	sequencing	

A11	Sterility testing			
1	Biological Indicator	No. of specimens received	Culture	x 1
2	Air Sampling	No. of specimens	Culture	x 1
	All Sampling	received	Biochemical	x 1
3	In-use Testing	No. of specimens	Culture	x 1
3	in-use resuity	received	Biochemical	x 1
4	Environment screen	No. of specimens	Culture	x 1
4		received	Biochemical	x 1
5	R.O water/ Endotoxin	No. of specimens	Culture	x 1
3		received	Biochemical	x 1
6	Starility Taction	No. of specimens	Culture	x 1
0	Sterility Testing	received	Biochemical	x 1
A12	Others (if test not listed)	No. of specimens received	Each indivdual test	x 1
	TOTAL		TOTAL	

B. MYCOLOGY

No.	Type of Tests No. Specimen		Tests Performed	No. Test performed		
			Direct Microscopy (KOH)	x 1		
B1	Microscopy	No. of specimens	Indian ink	x 1		
וט	Wilcroscopy	received	LPCB stain	x 1		
			Pneumocystis carinii IF	x 1		
			Plate culture	x 1		
			Slide culture	x 1		
		No of opposite and	Biochemical (M)	x 1		
B2	Culture and sensitivity	No. of specimens received	Biochemical(C)	x 1		
		received	Biochemical(A)	x 1		
			Sensitivity testing E-test	x 1		
			Sensitivity testing broth method	x 1		
В3	Serology					
1	Aspergillus galactomannan Ag	No. of specimens received for each	Aspergillus Galactomannan Ag	x 1		
2	Candida mannan Ag	test	Candida Mannan Ag	x 1		
3	Cryptococcal Ag	No. of specimens	Cryptococcal Ag (serum/CSF)	x 1		
?	(serum/CSF)	received	Cryptococcal Ag (serum/CSF) dilution	x 1		
4	Histoplasma IgM	No. of specimens received	Histoplasma IgM	x 1		
B4	PCR					
		No of opposite and	Manual PCR	x 1		
1	Fungal PCR	No. of specimens received	Semi-automated PCR	x 1		
		received	Fully automated PCR	x 1		
2	16s sequencing	No. of specimens	16s sequencing Forward sequencing	x 1		
	105 Sequencing	received	16s sequencing Reverse sequencing	x 1		
B5	Others (if test not listed)	No. of specimens received	Each indivdual test	x 1		
	TOTAL		TOTAL			

C. PARASITOLOGY

No.	Type of Tests	No. Specimens	Tests Performed	No. Test performed	
		No of oppointing	Microscopy	x 1	
1	Acanthamoeba/Naegleria	No. of specimens received	Culture	x 1	
		10001100	Multiplex PCR	x 1	
		No of anasimona	Microscopy	x 1	
2	Amoebiasis	No. of specimens received	Serology	x 1	
			PCR	x 1	
3	Cryptosporidium spp	No. of specimens	Modified DMSO stain	x 1	
4	Cysticercosis	received for each test	ELISA	x 1	
			Giemsa stain	x 1	
5	Filariasis	No. of specimens	PanLF assay	x 1	
J	i ilailasis	received	ICT Bancroftian assay	x 1	
			Manual/Semi automated PCR	x 1	
6	Giardia lamblia	No. of specimens	Microscopy	x 1	
<u> </u>	Giatula lattibila	received	Culture	x 1	
7	Helminth diagnosis	No. of specimen	Macroscopy	x 1	
,	r tollillitur diagridaia	received	Culture	x 1	
8	Hydatid disease	No. of specimens received	ELISA	x 1	
	Leishmaniasis	No. of specimens received	Giemsa stain	x 1	
9			ELISA	x 1	
		received	PCR	x 1	
	Malaria	No. of specimens received	Giemsa stain	x 1	
10			Rapid test	x 1	
10			IF Antibody test	x 1	
			Seminested PCR	x 1	
11	Microsporodium spp	No. of specimens	Gram chromotrope stain	x 1	
12	Schistosomiasis	received for each test	ELISA	x 1	
			Direct smear	x 1	
13	Stool Ova & Cyst	No. of specimens received	Fecal concentration	x 1	
	-	received	Trichrome stain	x 1	
14	Toxocariasis	No. of specimens received	ELISA	x 1	
			ELSA IgM	x 1	
		Marie Carri	ELISA IgG	x 1	
15	Toxoplasma	No. of specimens received	IFAT - Ig G	x 1	
13	ι ολυμιασιπα	16661160	IFAT - Ig M	x 1	
			IgG avidity	x 1	
			IgA	x 1	
16	Trichinellosis	No. of specimens received	ELISA	x 1	
17	Trypanosomiasis	No. of specimens received	Giemsa stain Manual/Semi automation PCR	x 1 x 1	
18	Others (if test not listed)	No. of specimens received	Each indivdual test	x 1	
	TOTAL		TOTAL		

D. IMMUNOLOGY

No.	Type of Tests	No. Specimens	Tests Performed	No. Test performed
1	Acetylcholine-receptor Antibody	No. of specimens received	Acetylcholine-receptor Antibody	x 1
		N/a af amaaimaana	ANA Colorzyme	x 1
2	ANA	No. of specimens received for each	ANA ELISA	x 1
2	AIVA	test	ANA IF	x 1
			ANA dilution	x 1
			ANCA ethanol IF	x 1
			ANCA formalin IF	x 1
3	ANCA		ANCA dot	x 1
			Myeloperoxidase ANCA	x 1
			PR3 ANCA	x 1
4	Anti-Aquaporin 4 (Anti-Aq4)		Anti-Aquaporin 4 (Anti-Aq4)	x 1
5	Anti-Beta 2 glycoprotein 1		Anti-Beta 2 glycoprotein 1 IgG	x 1
	Antibody		Anti- Beta 2 glycoprotein 1 lgM	x 1
6	Anti- cardiolipin Antibody		Anti- cardiolipin IgM	x 1
0			Anti- cardiolipin IgG	x 1
7	Anti-Cyclic Citrullinated Protein (CCP/ACPA)	No. of specimens received for each	Anti-Cyclic Citrullinated Protein (CCP/ACPA)	x 1
8	Anti-Deamidated-Gliadin	test	Anti-Deamidated-Gliadin IgA	x 1
0	Antibody		Anti-Deamidated-Gliadin IgG	x 1
9	Anti-desmoglein 1, Anti- desmoglein 3		Anti-desmoglein 1, Anti-desmoglein 3	x 1
10			Anti - dsDNA EIA	x 1
	Anti-dsDNA		Anti - dsDNA IF	x 1
			Anti - dsDNA Colorzyme	x 1
11	Anti- gastric parietal cell		Anti- gastric parietal cell	x 1
12	Anti-Glomerular Basement Membrane (GBM)		Anti-Glomerular Basement Membrane (GBM)	x 1
13	Anti-Liver-Kidney Microsomal (LKM)		Anti-Liver-Kidney Microsomal (LKM)	x 1
14	Anti-Mitochondrial Antibody (AMA)		Anti-Mitochondrial Antibody (AMA)	
15	Anti-N-Methyl-D-Aspartate Receptor (NMDAR)		Anti-N-Methyl-D-Aspartate Receptor	x 1
16	Anti- Smooth Muscle Antibody (ASMA)		Anti- Smooth Muscle Antibody (ASMA)	x 1
		No. of specimens	Anti-Tissue Transglutaminase (tTG) IgA	x 1
		received for each	Anti-Tissue Transglutaminase (tTG) IgG	x 1
17	Coeliac Antibodies	test	Anti-endomysium Ig A	x 1
17	Coellac Artibodies		Anti-endomysium IgG	x 1
			Anti-gliadin IgA	x 1
			Anti-gliadin IgG	x 1
18	Dihydrorhodamine assay (DHR)		Dihydrorhodamine assay (DHR)	
19	Eosinophilic Cationic Protein		Eosinophilic Cationic Protein	x 1
20	Extractable Nuclear Antigen (ENA) screening	No. of specimens received	Extractable Nuclear Antigen (ENA) screening	x 1
21	Extractable Nuclear Antigen (ENA) confirmation	No. of specimens received	Specific ENA antibodies	x no. of specific ENA tested
22	Gamma-aminobutyric acid-b Receptor (GABA) Antibody	No. of specimens received	Gamma-aminobutyric acid-b Receptor (GABA) Antibody	x 1

		No. of specimens	Anti-GM1, Anti-GM2, Anti-GM3, Anti-GD1a, Anti-GD1b, Anti-GT1b, Anti-GQ1b	x 1
23	Gangliosides Antibodies	received	Anti-GM1, Anti-GM2, Anti-GM3, Anti-GD1a, Anti-GD1b, Anti-GT1b, Anti-GQ1b	x 1
24	Human leukocyte antigens (HLA) Antibody Detection (Donor Specific Antibody)		Human leukocyte antigens (HLA) Antibody Detection (Donor Specific Antibody)	x 1
25	Human leukocyte antigens (HLA) Crossmatch (Complement Dependent Cytotoxicity)	No. of specimens received for each test	Human leukocyte antigens (HLA) Crossmatch (Complement Dependent Cytotoxicity)	x 1
26	Human leukocyte antigens (HLA) Crossmatch (Flow Cytometry)		Human leukocyte antigens (HLA) Crossmatch (Flow Cytometry)	x 1
	Human leukocyte antigens		Low / medium resolution (SSP/SSO)	x 1
27	(HLA) Typing Class I (Loci A, B and C)		High resolution	x molecular markers tested
	Human leukocyte antigens		Low / medium resolution (SSP/SSO)	x 1
28	(HLA) Typing Class I and II (Loci A, B and DR)	No. of specimens received for each	High resolution	x molecular markers tested
	Human leukocyte antigens	test	Low / medium resolution (SSP/SSO)	x 1
29	(HLA) Typing Class II (Loci DR and DQ)		High resolution	x molecular markers tested
30	Human leukocyte antigens (HLA) Typing for Disease Association		Human leukocyte antigens (HLA) Typing for Disease Association	x 1
31	IgE specific allergen	No. of specimens received	IgE specific allergen	x no of allergen tested
32	Leukocytes Adhesion Deficiency Type 1	No. of specimens received	Leukocytes Adhesion Deficiency Type 1	x 1
33	Liver autoantibodies - screening	No. of specimens received	Liver autoantibodies screening	x 1
34	Liver autoantibodies - Specific	No. of specimens received	Specific liver antibody	x specific liver antibodies tested
35	Lymphocytes proliferation test		Lymphocytes proliferation test	x 1
36	Phagocytic function test		Phagocytic function test	x 1
37	PNS Antibodies		PNS Antibodies - Anti-Hu, Anti-Ri, Anti- Ma, Anti-Yo, Amphiphysin, CV2	x 1
38	Rheumatoid factor (RF)	No. of specimens received for each test	RF isotype ie. lgM/lgG/lgA	x no. of isotype tested
39	Skin Antibodies - Anti BP 180, Anti BP 230		Skin Antibodies - Anti BP 180, Anti BP 230	x 1
40	Tryptase		Tryptase	x 1
41	T & B Lymphocyte Subset Enumeration (Dual platform method)		CD3/4/8/45/19/56/16	x MoAb used
42	Others (if test not listed)	No. of specimens received	Each indivdual test	x 1
	TOTAL		TOTAL	

E. VIROLOGY

No.	Type of Tests	No. Specimens	Tests Performed	No. Test performed
E1	Electron microscopy	No. of specimens received	Electron microscopy	x 1
E2	Antigen detection			
1	Rotavirus		Rotavirus	x 1
2	Norovirus	No. of specimens	Norovirus	x 1
3	Adenovirus	received for each tests	Adenovirus	x 1
4	Rabies virus		Rabies virus Ag IFAT	x 1
E3	SEROLOGY		The state of the s	
			Chikungunya IgM	x 1
1	Chikungunya Antibody		Chikungunya IgG	x 1
			CMV IgM	x 1
2	CMV Antibody		CMV IgG	x 1
3	Coxsackie virus		Coxsackie virus	x 1
	COXSACKIE VIIUS			
			Dengue Combo – NS1, IgM/IgG	x 1
4	Dengue Antigen/Antibody		Dengue IgG	x 1
			Dengue IgM	x 1
			Dengue NS I Antigen	x 1
5	EDV Antibody		EBV IgG	x 1
5	EBV Antibody		EBV IgM	x 1
6	Enteroviruses Antigen		Enteroviruses Ag	x 1
	3		Hantavirus Antibody PA	x 1
7	Hantavirus Antibody		Hantavirus IgG	x 1
	-		Hantavirus IgM	x 1
8	HAV Antibody		HAVAb Total	x 1
0	HAV Antibody		HAV IgM	x 1
9	HBc Antibody		HBc IgM	x 1
3	FIBC Antibody	No. of specimens	HBc Total Antibody	x 1
10	HBe Antibody	received for each test	Hbe Antibody	x 1
11	HBe Antigen	1621	Hbe Antigen	x 1
12	HBsAg		HBsAg	x 1
13	HBsAg Confirmatory		HBsAg Confirmatory	x 1
14	HBsAb		HBs Ab	x 1
15	HCV Ab		HCV Ab	x 1
16	HCV Confirmatory		HCV Confirmatory (LIA)	x 1
17	HCV Supplementary (PA)		HCV Supplementary (PA)	x 1
18	HDV Antibody		HDV IgM	x 1
10	HDV Antibody		HDV IgG	x 1
19	HEV Antibody		HEV IgM	x 1
19	TIEV Antibody		HEV IgG	x 1
20	HHV6 Antibody		HHV6 IgM	x 1
	THIVE MINDOUS		HHV6 IgG	x 1
			HIV Ab ELISA	x 1
21	HIV Antibody		HIV Ab PA	x 1
			HIV Ab western blot/LIA	x 1
22	HIV Ag		HIV Ag	x 1
23	HIV Ag/Ab		HIV Ag/Ab	x 1
24	HSV Antibody		HSV IgM	x 1
- '			HSV IgG	x 1

Respiratory viruses identification No. of specimens received Parainfluenza 2 Parainfluenza 3 Parainfluenza 5 Parainfluenza 2 Parainfluenza 3 Parainfluenza 2					
Antibody Measles Antibody No. of specimens received for each test Mespiratory viruses screening Respiratory viruses screening No. of specimens received for each test No. of specimens received for each fest No. of specimens received fest or each fest No. of specimens received ferth fest fest No. of specimens received ferth fest fest fest fest fest fest fest fest	25	HTVL Antibody		HTVL Ab	x 1
Antibody Measles Antibody Measles Antibody Measles IgM Murps IgG Respiratory viruses identification eg. Influenza A - Influenza A - Influenza B - Parainfluenza 1 - Parai	00	Japanese encephalitis		Japanese encephalitis IgM	x 1
Measies Antibody	26			Japanese encephalitis IgG	x 1
Mumps Antibody Respiratory viruses Res	07			Measles IgM	x 1
Mumps IgM Mump	27	Measles Antibody	No. of specimens	· · ·	
Mumps IgG National Parvovirus B 19 Antibody Respiratory viruses Screening Respiratory viruses Respiratory viruses Respiratory viruses Influenza B Infl				· · ·	
Parvovirus B 19 Antibody Parvovirus B 19 IgM Parvovirus B 10 Influenza B Parvirus B Influ	28	Mumps Antibody	test		
Parvovirus B 19 Antibody Respiratory viruses Sarcening Respiratory viruses Respiratory syncitial virus Respiratory viruses Parainfluenza P			-		
Respiratory viruses	29	Parvovirus B 19 Antibody		<u> </u>	
Respiratory viruses identification e.g. Influenza A - Influenza B - Parainfluenza 1 - Parainfluenza 2 - Parainfluenza 3 - Parainfluenza 2 - Parainfluenza 3 - Parainfluenza 2 - Parainfluenza 3 - Parainfluenza 3 - Parainfluenza 2 - Parainfluenza 3 - Parainfluenza 2	30		-		
Rubella Antibody No. of specimens received for each test VZV IgM X 1	31			eg Influenza A - Influenza B - Parainfluenza 1 - Parainfluenza 2 - Parainfluenza 3 - Adenovirus - Respiratory syncitial virus	,
SZ VZV Antibody Teceived for each test VZV IgM X 1		Duballa Antibadu		Rubella IgM	x 1
Section Sect	32	Rubella Antibody		Rubella IgG	x 1
E4 MOLECULAR 1 BK virus PCR 2 Chikugunya virus 3 CMV 4 Coronavirus 5 Coxsackie virus A16,A24 6 Coxsackie B 7 Crimerian Congo haemorrhagic fever received for each test 9 Dengue virus 10 Ebola virus 11 Enterovirus 71 12 HAV 13 HBV quantitation 14 HCV quantitation 15 HCV genotyping 16 HIV - Drug resistant testing 17 HIV genotyping Assay 18 HIV RNA (Paediatric) 19 HIV Viral Load 19 HIV Viral Load 10 HSV 1/2 21 Identification by 16sRNA 22 Japanese Encephalitis 23 JC Virus PCR BK virus PCR genome detection and quantitation 2 x 1 2 Chikugunya virus 2 No Cotspackie in A 1 2 Chikugunya virus 2 No. of specimens received for each test 2 Japanese Encephalitis 2 No. of specimens received for each test 3 HV Viral Load 4 V 1 4 Coronavirus 5 ChW genome detection and quantitation 7 Chikugunya virus 7 ChW genome detection and quantitation 7 Chikugunya virus 7 ChW genome detection and quantitation 7 Chikugunya virus 7 ChW genome detection and quantitation 7 Chikugunya virus 7 ChW genome detection and quantitation 7 Chikugunya virus 7 ChW genome detection and quantitation 7 Chikugunya virus 7 ChW genome detection and quantitation 7 A 1 7 HAV genotyping Asay 8 A 1 8 HIV - Drug resistant testing 9 Dengue virus serotyping 9 Dengue virus serotyp	00	00 1/71/ A : ('b : 1		VZV IgM	x 1
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Chikugunya virus Chikugunya virus X 1	E4	MOLECULAR			
2 Chikugunya virus 3 CMV 4 Coronavirus 5 Coxsackie virus A16,A24 6 Coxsackie B 7 Crimerian Congo haemorrhagic fever received for each 10 Ebola virus 10 Ebola virus 11 Enterovirus 71 11 Enterovirus 71 12 HAV 13 HBV quantitation 14 HCV quantitation 15 HCV genotyping 16 HIV - Drug resistant testing 17 HIV genotyping Assay 18 HIV RNA (Paediatric) 19 HIV Viral Load 20 HSV 1/2 21 Japanese Encephalitis 23 JC Virus PCR A Coronavirus Coronavirus A 1 Coronavirus Coxsackie virus A16,A24 A 1 Coxsackie B A 2 1 Coxsackie B A 3 1 Coxsackie Virus A16,A24 A 1 Dengue virus genome detection A 1 Ebola virus Bebola virus A 1 HAV A 1 HBV quantitation A 1 HCV quantitation A 1 HCV quantitation A 1 HIV - Drug resistant testing A 1 HIV genotyping assay A 1 HIV genotyping assay sequencing A 1 HIV RNA A 1 HIV RNA A 1 HIV RNA A 1 HIV RNA A 1 HIV Viral Load A 1 HIV Viral	1	1 BK virus BCB		BK virus PCR genome detection and	v 1
CMV					
4 Coronavirus 5 Coxsackie virus A16,A24 6 Coxsackie B 7 Crimerian Congo haemorrhagic fever 8 Dengue virus 9 Dengue virus serotyping 10 Ebola virus 11 Enterovirus 71 12 HAV 13 HBV quantitation 14 HCV quantitation 15 HCV genotyping 16 HIV - Drug resistant testing 17 HIV genotyping Assay 18 HIV RNA (Paediatric) 19 HIV Viral Load 20 HSV 1/2 21 Identification by 16sRNA 22 Japanese Encephalitis 23 JC Virus PCR No. of specimens received for each test Coronavirus 2 Coxsackie B 2 x 1 Coxsackie B 2 x 1 Coxsackie B 2 x 1 Coxsackie B 3 x 1 Dengue virus genome detection 3 x 1 Enterovirus 71 HAV 4 HAV 4 HAV 4 x 1 HAV 4 HBV quantitation 4 HCV quantitation 5 HCV genotyping 7 Thiv genotyping assay 8 x 1 Thiv genotyping assay 9 x 1 Thiv genotyping assay 1 HIV genotyping assay sequencing 1 HIV RNA 1 HIV Viral Load 1 HIV RNA 1 HIV Viral Load 2 x 1 Thiv RNA 3 Thiv RNA 4 Thiv Viral Load 5 Thiv RNA 7 Thiv RNA 7 Thiv RNA 7 Thiv RNA 8 Thiv RNA 8 Thiv RNA 8 Thiv RNA 8 Thiv RNA 9 T				Chikugunya virus	x 1
5 Coxsackie virus A16,A24 6 Coxsackie B 7 Crimerian Congo haemorrhagic fever 8 Dengue virus 9 Dengue virus serotyping 10 Ebola virus 11 Enterovirus 71 11 Enterovirus 71 11 HCV quantitation 14 HCV quantitation 15 HCV genotyping 16 HIV - Drug resistant testing 17 HIV genotyping Assay 18 HIV RNA (Paediatric) 19 HIV Viral Load 20 HSV 1/2 21 Identification by 16sRNA 22 Japanese Encephalitis 23 JC Virus PCR No. of specimens received for each test Coxsackie B		CMV		CMV genome detection and quantitation	x 1
6 Coxsackie B 7 Crimerian Congo haemorrhagic fever 8 Dengue virus 9 Dengue virus serotyping 10 Ebola virus 11 Enterovirus 71 12 HAV 13 HBV quantitation 14 HCV quantitation 15 HCV genotyping 16 HIV - Drug resistant testing 17 HIV genotyping Assay 18 HIV RNA (Paediatric) 19 HIV Viral Load 20 HSV 1/2 21 Identification by 16sRNA 22 Japanese Encephalitis 23 JC Virus PCR No. of specimens received for each test No. of specimens received for each test Coxsackie B Crimerian Congo haemorrhagic fever x 1 Crimerian Congo haemorrhagic fever x 1 Crimerian Congo haemorrhagic fever x 1 Dengue virus genome detection x 1 HAV HAV X 1 HAV HBV quantitation x 1 HCV quantitation x 1 HCV quantitation x 1 HIV genotyping assay x 1 HIV genotyping assay x 1 HIV genotyping assay sequencing x 1 HIV RNA HIV genotyping assay sequencing x 1 HIV RNA HIV genotyping assay sequencing x 1 HIV RNA HIV genotyping assay sequencing x 1 HIV RNA HIV genotyping assay sequencing x 1 HIV Genoty	4	Coronavirus		Coronavirus	x 1
7 Crimerian Congo haemorrhagic fever 8 Dengue virus 9 Dengue virus serotyping 10 Ebola virus 11 Enterovirus 71 12 HAV 13 HBV quantitation 14 HCV quantitation 15 HCV genotyping 16 HIV - Drug resistant testing 17 HIV genotyping Assay 18 HIV RNA (Paediatric) 19 HIV Viral Load 20 HSV 1/2 21 Identification by 16sRNA 22 Japanese Encephalitis 23 JC Virus PCR No. of specimens received for each test Crimerian Congo haemorrhagic fever x 1 Dengue virus genome detection x 1 Dengue virus serotyping 1 K 1 Dengue virus genome detection x 1 Dengue virus genome detection x 1 HAV HAV HAV HBV quantitation x 1 HIV quantitation x 1 HIV - Drug resistant testing X 1 HIV genotyping assay X 1 HIV genotyping assay X 1 HIV genotyping assay sequencing X 1 HIV RNA HIV Viral Load X 1 HIV RNA HIV Viral Load X 1 HIV Viral Load X	5	Coxsackie virus A16,A24		Coxsackie virus A16,A24	x 1
The state of the	6	Coxsackie B	1	Coxsackie B	x 1
9 Dengue virus serotyping 10 Ebola virus 11 Enterovirus 71 12 HAV 13 HBV quantitation 14 HCV quantitation 15 HCV genotyping 16 HIV - Drug resistant testing 17 HIV genotyping Assay 18 HIV RNA (Paediatric) 19 HIV Viral Load 20 HSV 1/2 21 Identification by 16sRNA 22 Japanese Encephalitis 23 JC Virus PCR Dengue virus genotine detection x 1 Dengue virus serotyping x 1 Enterovirus 71 x 1 HAV HBV quantitation x 1 HCV quantitation x 1 HCV genotyping x 1 HIV genotyping assay X 1 HIV genotyping assay X 1 HIV RNA HIV RNA HIV Viral Load X 1 HOW In Load X 1 HIV Viral Load X 1	7			Crimerian Congo haemorrhagic fever	x 1
10 Ebola virus 11 Enterovirus 71 12 HAV 13 HBV quantitation 14 HCV quantitation 15 HCV genotyping 16 HIV - Drug resistant testing 17 HIV genotyping Assay No. of specimens received 18 HIV RNA (Paediatric) 19 HIV Viral Load 20 HSV 1/2 21 Identification by 16sRNA 22 Japanese Encephalitis No. of specimens received for each test 10 Ebola virus Enterovirus 71 X 1 HAV X 1 HBV quantitation X 1 HCV quantitation X 1 HIV - Drug resistant testing X 1 HIV genotyping assay X 1 HIV genotyping assay sequencing X 1 HIV RNA HIV RNA X 1 HIV RNA X 1 HIV Viral Load	8	Dengue virus	test	Dengue virus genome detection	x 1
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12HAV13HBV quantitationHBV quantitation14HCV quantitationX 115HCV genotypingHCV genotyping16HIV - Drug resistant testingX 117HIV genotyping AssayNo. of specimens receivedHIV genotyping assayX 118HIV RNA (Paediatric)HIV genotyping assay sequencingX 119HIV Viral LoadHIV RNAX 120HSV 1/2HIV Viral LoadX 121Identification by 16sRNAHIV Viral LoadX 122Japanese EncephalitisX 123JC Virus PCRJC Virus PCR genome detection and quantitationX 1	10	Ebola virus		Ebola virus	x 1
HBV quantitation HCV quantitation HCV quantitation X 1	11	Enterovirus 71	1	Enterovirus 71	x 1
14 HCV quantitation15 HCV genotypingHCV genotypingX 116 HIV - Drug resistant testingHIV - Drug resistant testingX 117 HIV genotyping AssayNo. of specimens receivedHIV genotyping assayX 118 HIV RNA (Paediatric)HIV genotyping assay sequencingX 119 HIV Viral LoadHIV RNAX 120 HSV 1/2HIV Viral LoadX 121 Identification by 16sRNAHIV Viral LoadX 122 Japanese EncephalitisNo. of specimens received for each testIdentification by 16sRNAX 123 JC Virus PCRJapanese EncephalitisX 124 JC Virus PCR genome detection and quantitationX 1	12	HAV		HAV	x 1
15 HCV genotyping HCV genotyping X 1	13	HBV quantitation		HBV quantitation	x 1
16HIV - Drug resistant testingHIV - Drug resistant testingx 117HIV genotyping AssayNo. of specimens receivedHIV genotyping assayx 118HIV RNA (Paediatric)HIV genotyping assay sequencingx 119HIV Viral Loadx 120HSV 1/2HIV Viral Loadx 121Identification by 16sRNAHSV 1/2x 122Japanese EncephalitisJapanese Encephalitisx 123JC Virus PCRJapanese Encephalitisx 124JC Virus PCR genome detection and quantitationx 1	14	HCV quantitation		HCV quantitation	x 1
16HIV - Drug resistant testingHIV - Drug resistant testingx 117HIV genotyping AssayNo. of specimens receivedHIV genotyping assayx 118HIV RNA (Paediatric)HIV genotyping assay sequencingx 119HIV Viral Loadx 120HSV 1/2HIV Viral Loadx 121Identification by 16sRNAHSV 1/2x 122Japanese EncephalitisJapanese Encephalitisx 123JC Virus PCRJC Virus PCR genome detection and quantitationx 1	15	·]	·	x 1
17	16	HIV - Drug resistant testing		HIV - Drug resistant testing	x 1
18 HIV RNA (Paediatric) 19 HIV Viral Load 20 HSV 1/2 21 Identification by 16sRNA 22 Japanese Encephalitis 23 JC Virus PCR HIV genotyping assay sequencing x 1 HIV RNA x 1 HIV Viral Load x 1 HIV Viral Load x 1 HIV Viral Load x 1 HSV 1/2 x 1 Identification by 16sRNA x 1 Japanese Encephalitis x 1 JC Virus PCR genome detection and quantitation x 1	47		No. of specimens		x 1
19HIV Viral LoadX 120HSV 1/2No. of specimens received for each testHIV Viral LoadX 121Identification by 16sRNAHSV 1/2X 122Japanese EncephalitisJapanese EncephalitisX 123JC Virus PCRJC Virus PCR genome detection and quantitationX 1	17	mrv genotyping Assay		HIV genotyping assay sequencing	x 1
20HSV 1/2No. of specimens received for each testHSV 1/2x 121Identification by 16sRNAIdentification by 16sRNAx 122Japanese EncephalitisJapanese Encephalitisx 123JC Virus PCRJC Virus PCR genome detection and quantitationx 1	18	HIV RNA (Paediatric)		HIV RNA	x 1
21 Identification by 16sRNA	19	HIV Viral Load	1	HIV Viral Load	x 1
21 Identification by 16sRNA received for each Identification by 16sRNA x 1	20	HSV 1/2	No of specimens	HSV 1/2	x 1
22Japanese EncephalitisJapanese Encephalitisx 123JC Virus PCRJC Virus PCR genome detection and quantitationx 1	21	Identification by 16sRNA		Identification by 16sRNA	x 1
23 JC Virus PCR genome detection and quantitation x 1	22	Japanese Encephalitis		Japanese Encephalitis	x 1
	23	JC Virus PCR			x 1
	24	Lassa Virus	1	Lassa Virus	x 1

25	Marburg		Marburg	x 1	
26	Measles		Measles	x 1	
27	Nipah virus		Nipah virus	x 1	
28	PanEnterovirus		PanEnterovirus	x 1	
29	Parvovirus		Parvovirus	x 1	
30	Rabies		Rabies	x 1	
		No. of specimens	Respiratory viruses multiplex PCR	x 1	
		received	FluA	x 1	
			FluB	x 1	
			H1N1	x 1	
31	Respiratory viruses		MERS CoV	x 1	
			SARS Coronavirus	x 1	
			Specific Respiratory viruses other than listed above	x no. viral species tested	
32	Rift valley nucleic acid		Rift valley fever	x 1	
33	Rotavirus		Rotavirus	x 1	
34	Rubella		Rubella	x 1	
35	St Louis Encephalitis	No. of specimens	St Louis Encephalitis	x 1	
36	Varicella zoster Virus	received for each tests	Varicella zoster Virus	x 1	
37	West Nile virus	iesis	West Nile virus	x 1	
38	Yellow virus		Yellow virus	x 1	
39	Zika virus		Zika virus	x 1	
E 5	VIRAL ISOLATION				
1	Chikungunya		Chikungunya	x 1	
2	CMV		CMV	x 1	
3	Coronavirus		Coronavirus	x 1	
4	Coxsackie virus	-	Coxsackie virus	x 1	
5	Dengue virus		Dengue virus	x 1	
6	Enteroviruses		Enteroviruses		
7	Herpes Simplex Virus (HSV)		Herpes Simplex Virus (HSV)		
8	Japanese Encephalitis		Japanese Encephalitis		
9	Measles	No. of specimens	Measles	x 1 x 1	
10	Mumps	received for each test	Mumps	x 1	
11	Non-Poliovirus Virus	1031	Non-Poliovirus Virus	x 1	
12	Paramyxovirus		Paramyxovirus	x 1	
13	Poliovirus Virus		Poliovirus Viral isolation	x 1	
14	Poliovirus Environmental Surveillance		Poliovirus Environmental Surveillance	x 1	
15	Rabies		Rabies	x 1	
16	Rubella		Rubella	x 1	
17	SARS Coronavirus		SARS Coronavirus	x 1	
18	Respiratory viruses	No. of specimens received	Respiratory viruses eg. Influenza A Influenza B Parainfluenza 1 Parainfluenza 2 Parainfluenza 3 Adenovirus Respiratory syncitial virus Metapneumovirus	x no. viral species tested	
E6	Others (if test not listed)	No. of specimens received	Each indivdual test	x 1	
	TOTAL		TOTAL		

PREANALYTICAL WORKLOAD (FOR 'NON-TEST' TECHNICAL ACTIVITIES)

Activity	Workload
Media preparation	Volume in litres
TB EQA slides	Number of slides
IIP slides	Number of slides

PATHOLOGY SERVICES MINISTRY OF HEALTH, MALAYSIA

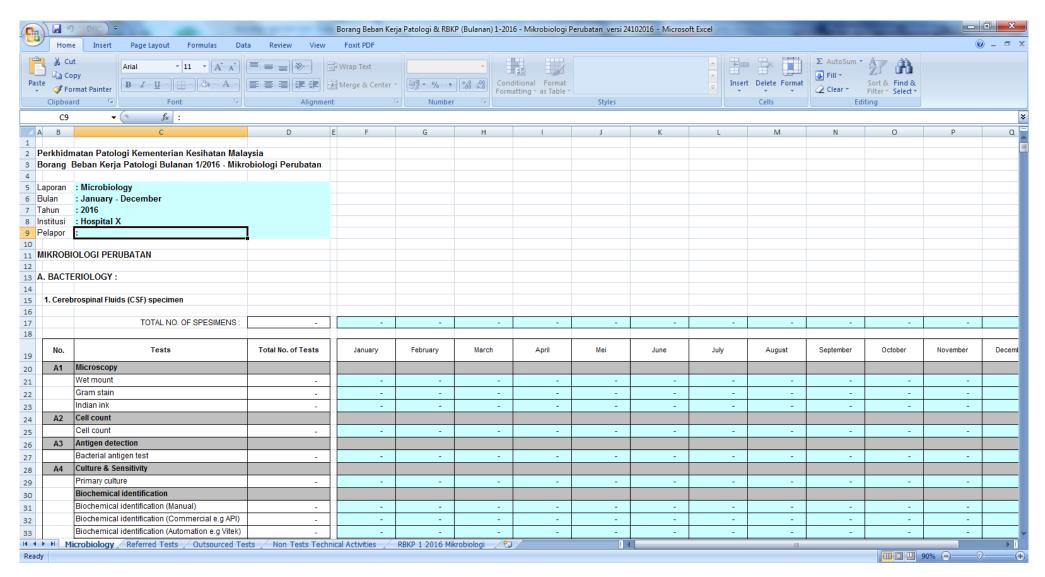
REPORT ON LABORATORY WORKLOAD: MEDICAL MICROBIOLOGY

FOR THE OF MONTH:	YEAR :

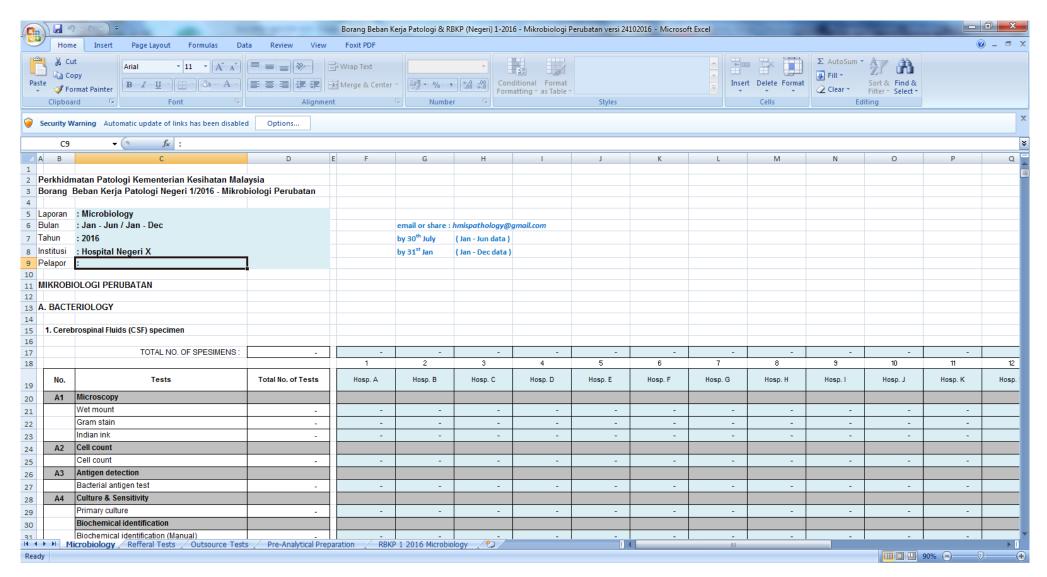
Month or Location of Hospitals,	Bacteriology		Mycology		Parasitology		Immunology		Virology		Total No.	Total No.
Institutions and Public Health Facilities	No. of specimens	No. of tests	of Specimens	of Tests								

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Borang Beban Kerja Patologi Negeri 1/2016 – Mikrobiologi Perubatan ('Excel Soft Copy')



7.5 GENETICS FOR CONGENITAL ANOMALY AND CANCER

7.5.1 Genetic testing for congenital anomalies and cancers are done in either dedicated/standalone molecular genetic laboratories or as part of pathology discipline, such as anatomic pathology. Therefore, workload is captured depending on where the tests are being performed.

7.5.2 Workload calculation and recording:

No.	Type of Tests		No. Specimens	Tests Performed	No. Test performed
1	Conventional karyotype		No. of specimens received	Chromosome Analysis	x 1
2	FISH Test				
	Syndrome	FISH Probes			x 1
2.1	Prader Willi / Angelman Syndrome	SNRPN (15q11.2)	No. of specimens received	Fluorescence- in-situ hybridization (FISH)	x 1
2.2	William Syndrome	Elastin(7q11.23)			x 1
2.3	DiGeorge Syndrome	N25(22q11.2)			x 1
2.4	Smith Magenis Syndrome	SMCR(11p11.2)			x 1
2.5	Miller Dieker Syndrome	MDS(11p13.3)			x 1
2.6	Wolf-Hirschhorn Syndrome	WHSCR(4p16.3)			x 1
2.7	Cri Du Chat Syndrome	5p15			x 1
2.8	Rubenstein Taybi	16p13.3			x 1
		SRY / CEP X			x 1
		ХрҮр			x 1
		WCP 1-22, X. Y			x 1
		p,q telomeric probe			x 1
		Centromeric probe			x 1
3	Karyolite Bobs Assay - Congenital anomalies (9 microdeletion & all chromosome subtelomeric)		No. of specimens received	BACs on beads technique	x 1
4	Duchenne Muscular Dystrophy / Becker Muscular Dystrophy			MLPA	x 1
5	Muenke Syndrome			PCR - RFLP & Sequencing	x 1
6	Rett Syndrome			DHPLC, MLPA & Sequencing	x 1
7	Array Comparative Genomic Hybridization (Oligo) - Congenital anomalies - Cancer genetics			Microarray analysis	x 1
8	Cancer Genetics : EGFR, BRAF, KRAS			Real time PCR	x 1
9	Cancer Genetics FISH test : ALK , ROS 1, N-MYC			FISH	x 1
10	Others (if test not listed)		No. of specimens received	Each indivdual test	x 1
			TOTAL		

8. CONCLUSION AND ACKNOWLEDGEMENT

It is hope that this guideline will help in the collection of standardized and meaningful national pathology data, thus aiding future planning, as well as, enhancing the cost effectiveness of services provided by the Ministry of Health laboratories.

The committee would like to thank all those who had contributed or involved in giving various input, directly or indirectly, towards the preparation of this guideline.

9. WORKING COMMITTEE AND CONTRIBUTORS

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